

Sexual Dichromatism Is Decoupled from Diversification over Deep Time in Fishes

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ABSTRACT: Sexually selected traits have long been thought to drive diversification, but support for this hypothesis has been persistently controversial. In fishes, sexually dimorphic coloration is associated with assortative mating and speciation among closely related species, as shown in classic studies. However, it is unclear whether these results can generalize to explain diversity patterns across ray-finned fishes, which contain the majority of vertebrate species and 96% of fishes. Here, we use phylogenetic approaches to test for an association between sexual dichromatism and diversification rates (speciation minus extinction) in ray-finned fishes. We assembled dichromatism data for 10,898 species, a data set of unprecedented size. We found no difference in diversification rates between monochromatic and dichromatic species when including all ray-finned fishes. However, at lower phylogenetic scales (within orders and families), some intermediate-sized clades did show an effect of dichromatism on diversification. Surprisingly, dichromatism could significantly increase or decrease diversification rates. Moreover, we found no effect in many of the clades initially used to link dichromatism to speciation in fishes (e.g., cichlids) or an effect only at shallow scales (within subclades). Overall, we show how the effects of dichromatism on diversification are highly variable in direction and restricted to certain clades and phylogenetic scales.

Keywords: diversification, fishes, sexual selection, speciation, color, phylogenetic scale.

Introduction

A major goal in evolutionary biology is to explain why some clades have more species than others (Wiens 2017). Fundamentally, new species are added to a clade through speciation and removed through extinction. Therefore, an important explanation for variation in richness among clades is that they differ in their net diversification rates (speciation minus extinction rates).

Diversification rates can be influenced by organismal traits that affect the probability of speciation or extinction. Darwin (1871) first observed that many of the most obvious differences between closely related species are in traits involved in courtship (Ritchie 2007), including the bright colors in males of many species (fig. 1). This observation led to the influential hypothesis that sexually selected traits play a role in the origin of new species (Boughman 2014). Classic papers proposed that sexual selection drives speciation through assortative mating (Lande 1981; West-Eberhard 1983). However, several additional mechanisms could also help explain how sexual selection could increase net diversification rates (Tsuji and Fukami 2020). For example, sexual selection may drive speciation through postzygotic isolation (Irwin 2020) or by facilitating ecological divergence (Albert et al. 2008). Sexual selection can also reduce extinction by helping populations adapt to changing environments (Lorch et al. 2003) or purge mutation loads (Lumley et al. 2015).

Conversely, sexual selection can also decrease net diversification rates. Sexual selection can inhibit speciation through negative assortative mating (Servedio and Burger 2014) or when sexes compete for resources (Bolnick and Doebeli 2003). If sexually dimorphic traits increase individual mortality, then sexual selection can drive species to extinction (Promislow et al. 1992; Martins et al. 2018). In sum, sexual selection may increase or decrease net diversification rates of clades through several mechanisms that act on speciation or extinction.

Comparative studies testing for an association between sexually selected traits and diversification rates have found mixed results. Sexual dichromatism was first found to be positively related to species richness and diversification on the basis of comparisons of sister clades of passerine birds (Barraclough et al. 1995). However, subsequent studies in birds found no such relationship (Morrow et al. 2003; Huang and Rabosky 2014). Studies in other animal clades are similarly divided on whether sexual selection increases, decreases,

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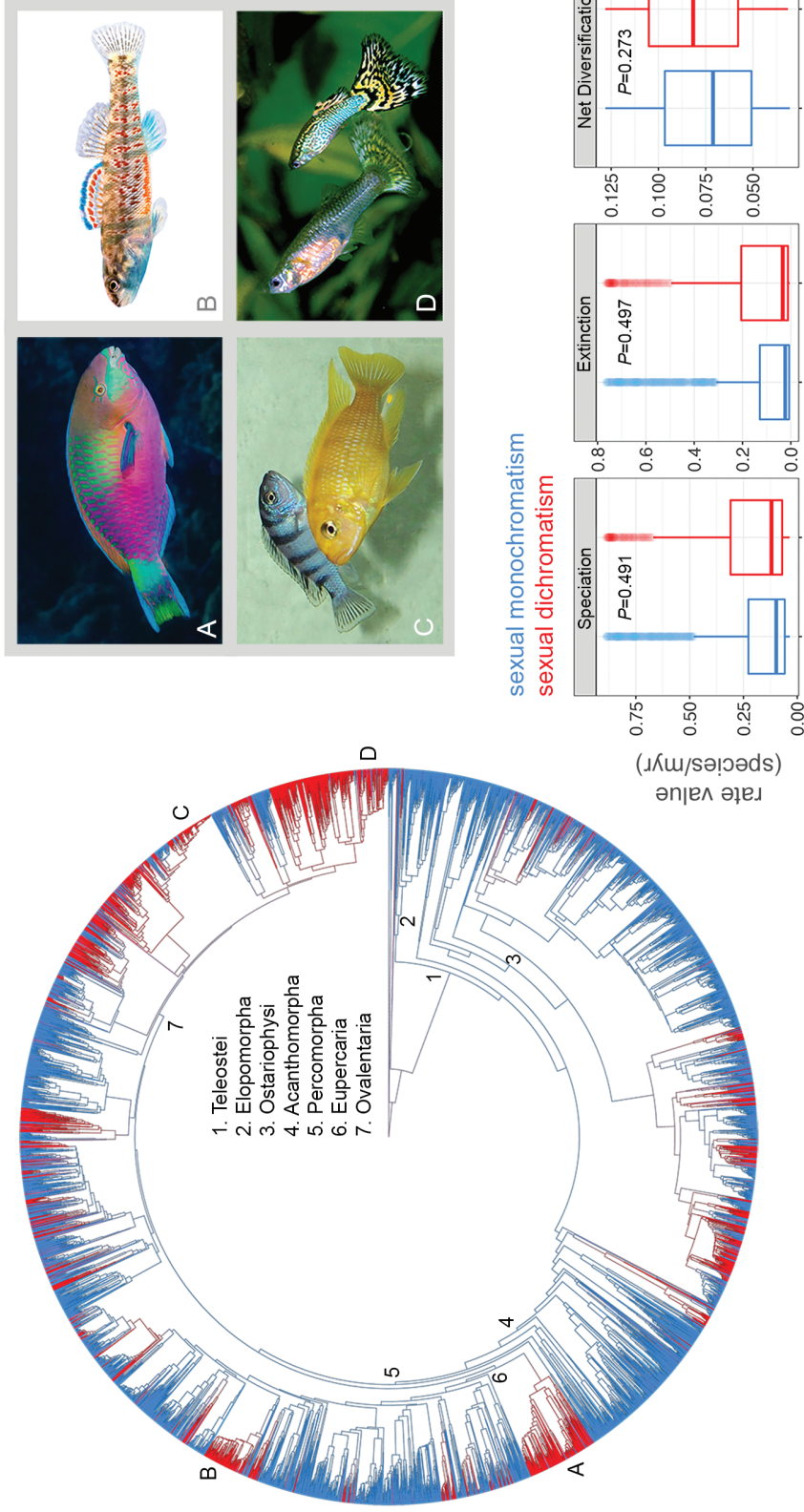


Figure 1: Sexual dichromatism and diversification across ray-finned fishes. An ancestral reconstruction of sexual dichromatism was estimated using the best-fit hidden state speciation and extinction model (table A14) on a phylogeny of all ray-finned fishes (Rabosky et al. 2018). Representative clades with prior work on sexual dichromatism and assortative mating, reproductive isolation, and/or diversification are shown in photographs: parrotfishes (Kazancıoğlu et al. 2009; A), darters (Mendelson 2003; B), cichlids (Seehausen et al. 1997; C), and guppies (Furness et al. 2019; D). Across ray-finned fishes, there were no significant differences in rates between sexually monochromatic and dichromatic species (table A15). Rates for monochromatic species are shown on the left in blue; rates for dichromatic species are shown on the right in red. Boxplots summarize rates for individual species, including medians, first and third quartiles, and outliers (outside the 95% confidence interval). Photograph credits: François Libert (A); Daniel MacGuigan (B); Wikimedia user Ged-commonswiki, public domain (C); Wikimedia user Marrabbio2, CC BY-SA 3.0 (D).

or has no effect on diversification (Kraaijeveld et al. 2011; Tsuji and Fukami 2020).

One possible explanation for these conflicting results is the different taxonomic scales among studies. A meta-analysis (Kraaijeveld et al. 2011) posited that studies at smaller scales (e.g., studies within bird genera) were more likely to recover a positive effect of sexual selection on diversification than studies of older clades (bird families). These authors suggested that this occurs because extinction prunes many of the species gained by speciation through sexual selection, so these species are not observed. A study that attempted to parse out the effects of taxonomic scale found no relationship between sexual dichromatism and speciation rates across all birds and none within most families (Huang and Rabosky 2014). Furthermore, a subsequent study in birds compared diversification rates among recently diverged sister species pairs and still found no relationship between dichromatism and speciation rates (Cooney et al. 2017). Thus, empirical evidence for this scaling hypothesis remains limited. However, it has been tested only in birds so far.

Despite uncertainty in other groups, current evidence for a positive effect of sexual dichromatism on speciation in ray-finned fishes is compelling (Kraaijeveld et al. 2011). With ~31,000 species, ray-finned fishes (Actinopterygii) contain ~96% of all fish species and ~50% of all vertebrates (Helfman et al. 2009). There is strong evidence that species-specific, sexually dichromatic colors are under sexual selection in fishes (Kodric-Brown 1990, 1998). In addition, male colors have been linked to assortative mating and/or reproductive isolation in several clades, including classic studies in cichlids (Seehausen et al. 1997, 2008; Elmer et al. 2009), darters (Mendelson 2003; Martin and Mendelson 2014), guppies (Houde and Endler 1990), and sticklebacks (Boughman 2001). Furthermore, sexual dichromatism has been linked with diversification over macroevolutionary timescales in cichlids (Wagner et al. 2012; McGee et al. 2020), parrotfishes (Kazancıoğlu et al. 2009; Choat et al. 2012), poeciliids (guppies and allies; Furness et al. 2019), and sunfishes (Smith et al. 2015). Most notably, sexual dichromatism helps explain why African cichlids have the fastest diversification rates among all animals (McGee et al. 2020). If the factors that promote reproductive isolation among populations also explain diversification rates over deeper timescales, then sexual dichromatism could help explain the striking differences in species diversity among ray-finned fish clades. For example, actinopterygian orders range from one to >4,000 species (table A1; tables A1–A28 are available online; Rabosky et al. 2018; Froese and Pauly 2019).

Here, we investigate the effect of sexual dichromatism on diversification rates across ray-finned fishes. This goal was shared by two previous studies that used sister clade comparisons to address diversification (Mesnick 1996; Mank

2007). Both studies found that groups with a greater prevalence of sexual dimorphism (in color and other features) were generally more species rich, which is consistent with faster diversification. However, the sister clade approach necessarily limits comparisons to only those groups that vary in the presence of dimorphism. Collectively, these two studies included 30.9% of ray-finned fish orders and 11.6% of families (Froese and Pauly 2019). This limitation may bias the range of diversification rates seen among monomorphic groups (e.g., ascertainment bias; Beaulieu and O'Meara 2018), potentially leading to incorrect inferences about the influence of sexual selection on diversification overall.

We improved on previous analyses of ray-finned fishes in three ways. First, we collected data on sexual dichromatism for 10,898 species, including 35% of described species, 89% of families, and 100% of orders (following taxonomy of Rabosky et al. 2018). Our data set is the largest on sexual dichromatism compiled for any animal group. Second, we used methods that separately estimate rates of speciation, extinction, and net diversification rather than using species richness as a proxy for diversification. Third, we performed analyses at three scales: across all ray-finned fishes, within clades in which sexual dichromatism evolved, and within smaller subsets of these clades. We also perform analyses that compare rank-free clades for a range of clade ages. By sampling broadly across taxa and scales, we can identify the scales at which sexual dichromatism drives diversification, if at all.

Material and Methods

Literature Search

We conducted an extensive literature search on the presence or absence of sexual dichromatism in actinopterygian fishes. We targeted information for those species that were represented in the phylogeny used (Rabosky et al. 2018; text A1; table A1; texts A1–A5 are available online). Our search was conducted from November 1, 2017, to June 1, 2019. We used the package *rfishbase* v.2.99 (Boettiger et al. 2012) to reconcile synonyms across literature sources. We focused on color differences over other forms of sexual dimorphism because these differences are (1) relatively straightforward to identify, (2) often reported when they exist (see below), and (3) linked to assortative mating, reproductive isolation, and/or diversification in many previous studies (see above).

Sexual dichromatism was defined here as any reported difference in color between the sexes at any stage in the adult life cycle, including differences in color pattern or intensity. This definition includes permanent and temporary dichromatism (Kodric-Brown 1998) as well as color changes

associated with sex change. This definition is agnostic regarding the degree of difference between the sexes, the mechanistic basis for color (e.g., melanic, carotenoid), and whether males or females are more colorful. We chose this definition for two reasons. First, our definition must apply across fishes even to clades that show only subtle dichromatism. Second, we do not always know which cases of dichromatism are unimportant for mate choice, as most fishes have not been studied behaviorally (Amundsen 2003).

We coded sexual dichromatism as present in a species if at least one source described sex differences in color (table A2). There were two ways that species could be coded as monochromatic (table A2). The first way was if a source stated that the species (or higher taxon to which it belonged) was monochromatic. However, we anticipated that many sources were unlikely to explicitly state that a species was monomorphic if the authors' goal was species identification (as for field guides). Therefore, we also coded species as monochromatic if the species' color in life was described but no sex differences were mentioned. The benefit of including these species is the increased power to detect variation in diversification rates (Davis et al. 2013). Our literature search supported our supposition that dichromatism is likely to be reported if it exists, but authors explicitly report monochromatism less often (text A2; table A3). Nevertheless, we performed analyses without these species and obtained similar results (see below; table A4).

Phylogenetic Framework

We used a time-calibrated maximum likelihood phylogeny (Rabosky et al. 2018) of ray-finned fishes including 11,638 species with DNA sequence data (36.9% of all described species). We included only species with sequence data because semirandom grafting of unsampled species is inappropriate for analyzing trait evolution at the species level (Rabosky 2015). We removed duplicate and unidentified species as well as species for which we could not find information on sexual dichromatism from the literature (see "Results"). Species sampling among orders in this reduced phylogeny (10,898 species) was correlated with their described richness ($n = 68$ orders; $r = 0.97$; $P < .001$; text A1).

We initially performed analyses at three phylogenetic scales: (1) across all ray-finned fishes, (2) within each of 18 clades individually, and (3) within eight selected subclades of the six largest of those 18 clades. Details for all 18 clades and eight subclades are given in table A5. To delimit clades in all cases, we targeted monophyletic higher taxa (families and orders) in which both monochromatism and dichromatism were present in $\geq 10\%$ of the species. State-dependent speciation and extinction (SSE) methods can give problematic results when one state is rarer than

10% (Davis et al. 2013). To maximize species sampling, we used orders instead of families in cases where we could include additional species while maintaining this proportion. These 18 clades contained 93–1,098 species in the tree (mean = 349.2, SD = 265.4). The six largest clades (Characiformes, Cichlidae, Cyprinidae, Cyprinodontiformes, Gobiiformes, and Scorpaeniformes) included eight families and subfamilies that were also suitable for analyses under these criteria. These eight subclades included 111–438 species (mean = 273.8, SD = 123.8).

To assess how phylogenetic uncertainty may affect our results, we repeated hidden state speciation and extinction (HiSSE) analyses (see below) on three clades with alternative published phylogenies: Cichlidae (Burruss and Tan 2017), Labridae (Siqueira et al. 2016), and Poeciliidae (Reznick et al. 2017). These clades were particularly important because of previous studies linking dichromatism and diversification within them (e.g., Kazancıoğlu et al. 2009; Wagner et al. 2012; Furness et al. 2019). Furthermore, species sampling in these trees was similar to that in the main tree (Rabosky et al. 2018). Thus, any differences in results should be related to the trees (topology and divergence times) and not sampling. Details of these alternative trees are in text A3 and table A6.

We also used a rank-free approach to select clades (Poe et al. 2020). To test whether temporal scale influences our results, we sliced the phylogeny at different time points. We collected all subtrees produced by slicing the tree at 100, 80, 60, and 40 million years (myr) ago, respectively. Slicing was performed using the *chainsaw2* function in BioGeoBEARS version 1.1.2 (Matzke 2014). We then filtered the resulting clades to find those suitable for SSE analyses (>100 species, both states present in $\geq 10\%$ of species). This yielded 12 clades ~ 100 myr in age, 16 at ~ 80 myr, 15 at ~ 60 myr, and six at ~ 40 myr, for a total of 49 clades. For a given slice, most clades were subsets of those from older slices. Details of rank-free clades are in tables A7–A10.

Diversification Analyses

We primarily tested for associations between sexual dichromatism and diversification using the HiSSE framework implemented in the R package *hisse* version 1.9.0 (Beaulieu and O'Meara 2016). The HiSSE framework is an advance over previous SSE methods because it can detect rate shifts that are not caused by the trait of interest. It can thereby reduce false associations between diversification rates and the focal trait (Rabosky and Goldberg 2015). We performed identical HiSSE analyses on each of the 27 clades separately (all ray-finned fishes, 18 fish clades, and eight subclades) and the 49 clades from the rank-free approach. We compared the fit of 14 dichromatism-dependent models of diversification (in which dichromatism accounts

for some or all of the diversification rate heterogeneity in the tree) to nine dichromatism-independent models (in which dichromatism is unrelated to diversification rate heterogeneity). We compared the fit of dichromatism-dependent models to dichromatism-independent models of similar complexity (Beaulieu and O'Meara 2016; Harrington and Reeder 2017). Details of the 23 models are given in table A11. To correct for incomplete species sampling, we calculated the proportion of unsampled species for each clade using rfishbase (Boettiger et al. 2012), assuming that this proportion was identical between monochromatic and dichromatic species. We did this because the total number of monochromatic and dichromatic species among ray-finned fishes is unknown. However, supplemental analyses suggest that monochromatic and dichromatic species have similar chances of being sampled in the phylogeny (text A1). Relative fit of models was determined using Akaike weights (AICw; Burnham and Anderson 2002).

When a dichromatism-dependent model of diversification had the best fit, a range of relationships between dichromatism and diversification were possible. Dichromatism could have positive or negative effects on speciation, extinction, and diversification. To characterize this relationship for each clade, we obtained model-averaged rates associated with each species, with the contribution of each model toward the mean proportional to that model's AICw (Caetano et al. 2018). Only models with $\geq 5\%$ of the AICw of the model set were included in model averaging to avoid undue influence of poorly supported models (Caetano et al. 2018). Next, we tested whether model-averaged speciation, extinction, and net diversification rates were significantly different between monochromatic and dichromatic species, using phylogenetic ANOVA (Garland et al. 1993) implemented in phytools version 0.6-44 (Revell 2012). Note that in some cases, a dichromatism-independent model received 100% of the AICw but mean rates were still significantly different between monochromatic and dichromatic species using phylogenetic ANOVA. We interpret this result as indicating that other (hidden) factors are ultimately responsible for rate shifts (Beaulieu and O'Meara 2016). A recent study suggested that it can be mathematically difficult to separate the contributions of speciation and extinction to changes in diversification through time (Louca and Pennell 2020), but it is unclear whether this problem applies to SSE models because they are more complex than birth-death models.

At the scale of all ray-finned fishes, we also tested the robustness of our HiSSE results using Bayesian analysis of macroevolutionary mixtures (BAMM). BAMM (Rabosky 2014; Rabosky et al. 2014) is a Bayesian framework for detecting shifts in diversification. There are two relevant differences between this approach and HiSSE. First, BAMM estimates diversification rate shifts independently from trait

evolution. Second, HiSSE is limited by the number of rate shifts (i.e., hidden states) it can model, while BAMM has no limit on potential rate shifts. We acknowledge that BAMM may still be relatively insensitive to rate variation, especially for small clades (Moore et al. 2016; Rabosky et al. 2017; Meyer and Wiens 2018). We used structured rate permutations on phylogenies (STRAPP; Rabosky and Huang 2016), which uses a permutation test to detect an association with a binary trait and diversification rate shifts estimated by BAMM. This approach is best applied only to very large phylogenies because many rate shifts are needed to gain enough power to detect associations with a trait (Rabosky and Huang 2016).

We processed the BAMM output performed by Rabosky et al. (2018) using BAMMtools version 2.1.6 (Rabosky et al. 2014). We generated a null distribution of the Mann-Whitney U -test statistic by performing 1,000 permutations of the BAMM rate shifts associated with each species, using the traitDependentBAMM function in BAMMtools. This null distribution was used to test for associations between dichromatism and rates of speciation, extinction, and diversification. We used alternative rates estimated using a time-constant and time-variable BAMM model. We also performed tests using only species with direct information about the presence or absence of dichromatism ($n = 8,613$ species).

Alternative Perspectives on Color

We tested the sensitivity of our results to (1) treating sexual dichromatism as a binary variable and (2) using human perceptions of color. These analyses are summarized here, with details in texts A4 and A5.

Darwin (1871) originally considered the degree of dimorphism to be a proxy for the strength of sexual selection. Therefore, even if sex differences are present, diversification rates may be elevated only when dichromatism is dramatic. To test this possibility, we used color data from the literature for two well-studied clades: Etheostominae (darters; Bossu and Near 2015) and Labridae (wrasses and parrotfishes; Hodge et al. 2020). These studies each quantified sexual dichromatism on a continuous scale using photographs of males and females of each species. We tested for a relationship between the degree of dichromatism and tip-associated speciation rates from BAMM (calculated by Rabosky et al. 2018) using phylogenetic generalized least squares (Freckleton et al. 2002). This relationship was not significant in either clade (table A12). High speciation rates were found along the continuum from monochromatic to extremely dichromatic species (fig. A1; figs. A1–A6 are available online). These results were consistent with those using binary state models for these two clades (see “Results”; text A4).

Like many previous studies (e.g., Wagner et al. 2012), our data set was based on human perceptions of fish color. Many birds that appear sexually monochromatic to humans are actually dichromatic in the ultraviolet (UV) spectrum (Eaton 2005). We are not aware of any studies in fishes that found that a species that was sexually dichromatic in the UV spectrum appeared monochromatic to humans, as is the case for birds (Eaton 2005; but see Siebeck et al. 2010 on UV patterns and species discrimination in fishes). Furthermore, many fishes that are capable of UV vision lack body coloration visible in UV (Siebeck and Marshall 2001). Fishes may use UV vision in many contexts, such as feeding (Losey et al. 1999). Thus, UV vision in fishes need not be relevant to sexual selection.

Nevertheless, we assessed how our results would be affected by relying on human color perception to code the presence of dichromatism (text A5). We used a compendium of λ_{\max} (peak sensitivity wavelength of photoreceptors) estimates across 277 species of ray-finned fishes (Schweikert et al. 2018) to identify fishes with UV-sensitive vision. First, we found that fishes with UV vision were significantly more likely to appear sexually dichromatic to humans than other fishes (χ^2 test: $\chi^2 = 5.97$, $P = .0145$, $n = 237$ species). This suggests that fishes with the greatest potential to see colors in UV are also likely to have dichromatic coloration detectable by humans. Second, we tested whether speciation rates were significantly different among monochromatic and dichromatic fishes, assuming an extreme case in which all fishes with UV vision are sexually dichromatic even if they were assigned as monochromatic based on human color. We found that any differences in speciation rates associated with dichromatism were driven by biased sampling of dichromatic cichlids in the vision data set. Once cichlids were removed, there were no differences in speciation rates between monochromatic and dichromatic species, regardless of whether we recoded species with UV vision as dichromatic (table A13). Overall, we found no evidence that our results across fishes would be overturned by the failure to observe sexual dichromatism outside the human visible spectrum.

Results

Prevalence of Sexual Dichromatism

We obtained information from the literature on the presence and absence of sexual dichromatism for 10,898 species of ray-finned fishes, including most (93.6%) of the species sampled in the phylogeny (Rabosky et al. 2018) and 34.6% of all described species. Data were obtained from 196 sources (data set S1, deposited in the Dryad Digital Repository [<https://doi.org/10.5061/dryad.hqbkzkh1bs>; Miller et al. 2021]), including primary literature, online databases, field guides, taxonomic compilations, aquarist resources,

and expert opinion (for two families with limited information). There was a mean of 2.5 sources associated with each species ($SD \pm 1.7$, range = 1–15).

Sexual dichromatism was reported in 3,154 sampled species (28.9% of species in our data set). This proportion is not significantly different from an earlier estimate of 27.0% (Mank 2007; χ^2 test: $\chi^2 = 1.45$, $P = .23$; text A1). This implies that the prevalence of sexual dichromatism would not change greatly with additional sampling. There was no correlation between the proportion of species sampled in the phylogeny for each order and the proportion of dichromatic species in that order ($n = 68$ orders; $r = 0.17$; $P = .16$; text A1). Thus, sexual dichromatism did not appear to influence species sampling in this phylogeny.

Diversification Rates across All Ray-Finned Fishes

Our results show that sexual dichromatism does not significantly impact diversification rates when all ray-finned fishes are included (fig. 1). Using HiSSE, a dichromatism-independent model received 100% of the AICw (table A14). Using phylogenetic ANOVA on model-averaged rates (table A15), we found no significant difference in speciation rates between monochromatic and dichromatic species (mean $\lambda_{\text{monochromatic}} = 0.197$ lineages per myr [0.058–0.227; first and third quantile]; $\lambda_{\text{dichromatic}} = 0.241$ [0.070–0.310]; $P = .491$). We also found no significant difference in extinction rates ($\mu_{\text{monochromatic}} = 0.123$ [0.007–0.128]; $\mu_{\text{dichromatic}} = 0.159$ [0.012–0.205]; $P = .497$) or net diversification rates ($r_{\text{monochromatic}} = 0.074$ [0.051–0.097]; $r_{\text{dichromatic}} = 0.081$ [0.058–0.105]; $P = .273$). Although SSE methods have the power to detect trait-dependent diversification in clades with only a few hundred species (Davis et al. 2013), we did not find such an association with 10,898 species.

Using STRAPP, we also found no significant differences between monochromatic and dichromatic species in rates of speciation ($P = .53$ under a BAMM time-variable model, $n = 10,898$ species), extinction ($P = .61$), or net diversification ($P = .53$), regardless of the BAMM model used (table A4). We also performed STRAPP analyses that excluded the 2,285 species that were coded as monochromatic based solely on color descriptions that did not mention differences between the sexes. Again, we found no differences in rates (table A4).

Patterns within Individual Clades

We first fit HiSSE models on 18 clades individually. For illustrative purposes, we compared the summed AICw of 14 dichromatism-dependent models with the summed AICw for nine dichromatism-independent models (table A11). Scores of individual models are given in table A14, and model-averaged rates are given in table A15. Dichromatism-dependent

models of diversification received >80% of the AICw in only four clades (figs. 2, A2; table A14). In Tetraodontiformes (pufferfishes and relatives), dichromatic species had higher speciation rates but also higher extinction rates, such that monochromatic species had higher net diversification rates. Centrarchiformes (sunfishes and relatives) showed no significant difference in speciation or diversification rates. However, extinction rates were significantly lower in dichromatic species. Anabantiformes (bettas and relatives) and Blenniiformes (blennies) had significantly positive relationships between dichromatism and net diversification. However, in Anabantiformes, the difference in diversification rates was again due to lower extinction rates in dichromatic species, not faster speciation rates.

Among the remaining 14 clades, seven showed no evidence for an effect of dichromatism on diversification (figs. 2, A3). The other seven clades had only equivocal evidence for a positive relationship (three clades; fig. A4) or a negative relationship (four clades; fig. A5). These seven equivocal cases occurred when dichromatism-dependent and dichromatism-independent models received similar support (e.g., Gobiiformes and Percidae; AICw of dichromatism-independent models = 40% and 58%, respectively) or when a dichromatism-independent model had strong fit but mean rates were still significantly different using phylogenetic ANOVA (table A15). This latter pattern implies that diversification is associated with other (hidden) factors that partially overlap with dichromatism (Beaulieu and O'Meara 2016).

Surprisingly, the 14 clades in which dichromatism-dependent models received equivocal or no support encompassed many classic model systems that linked sexual dichromatism to reproductive isolation and/or speciation. These clades included Cichlidae (sum of AICw of dichromatism-independent models = 100%; $n = 720$ species), Cyprinodontiformes (guppies and relatives; AICw = 100%; $n = 572$), Labridae (wrasses and parrotfishes; AICw = 100%; $n = 337$), and Percidae (darters; AICw = 58%; $n = 218$). We found similar results when using alternative phylogenies of Cichlidae and Labridae (tables A16, A17).

Patterns at Shallower Phylogenetic Scales

Among the 18 clades, the six largest were Characiformes, Cichlidae, Cyprinidae, Cyprinodontiformes, Gobiiformes, and Scorpaeniformes. Dichromatism-independent models received 100% of the AICw in five of these six clades (fig. 2; table A14). However, support for dichromatism-dependent models strongly increased in four of eight subclades, relative to the larger clade to which they belonged (to 39% of the AICw in Characidae, 65% in Poeciliidae, 86% in Leuciscinae, and 100% in Gobiidae; fig. 3; table A18). The relationships between dichromatism and diversification were

variable among these groups. Characidae (tetras) and Leuciscinae (minnows) had higher net diversification rates in monochromatic species (table A19; fig. A6). Dichromatic gobies (Gobiidae) had higher speciation rates but also higher extinction rates, leaving no significant difference in net diversification rates. In Poeciliidae, higher net diversification rates associated with dichromatism were due to lower extinction rates, not higher speciation rates (table A19; fig. A6). We also recovered a positive effect of dichromatism on net diversification using an alternative phylogeny of Poeciliidae (Reznick et al. 2017). This effect was due to both speciation and extinction rates using this tree (tables A16, A17). We found no evidence for higher diversification rates associated with dichromatism in the subfamilies Pseudocrenilabrinae (African cichlids) and Cichlinae (Neotropical cichlids; fig. 3; table A18; fig. A6), mirroring the results for Cichlidae as a whole (fig. 2). This surprising result is robust to use of an alternative cichlid phylogeny (Burruss and Tan 2017; tables A16, A17).

Rank-Free Time Slice Approach

HiSSE results among 49 clades selected using time slices (fig. 4A; tables A7–A10) showed patterns similar to results from ranked clades. Only one of the 12 oldest clades (100-my time slice) had strong support for any effect of sexual dichromatism on diversification (fig. 4B; table A20). In this clade (loaches), the summed AICw of dichromatism-dependent models was 85%, and dichromatism had a negative effect on net diversification (table A21). The corresponding weight for the remaining 11 clades ranged from 0% to 41% and was <5% for eight clades (table A20).

Support for dichromatism-dependent models was higher among the clades from the 80-, 60-, or 40-my time slices (fig. 4B; tables A22–A27). Most of these clades were nested within the 12 oldest clades from the 100-my time slice (tables A7–A10). Among these 12 clades, eight contained subclades with stronger support for dichromatism-dependent models than the clade as a whole (fig. 4B). For example, cichlids finally showed the expected support for dichromatism-dependent diversification at the youngest level. Subclades within the African and Neotropical cichlid lineages from the 40-my time slice each had strong support for dichromatism-dependent models (100% and 91% of the AICw, respectively; table A26), with a positive effect of dichromatism on net diversification (fig. 4B; table A27). Among all clades with strong support for dichromatism-dependent models, there was no common tendency for dichromatism to increase or decrease speciation, extinction, or net diversification.

Support for dichromatism-dependent HiSSE models did not linearly increase with decreasing crown-group age (fig. 4C). The 100-my subset of clades collectively had poor

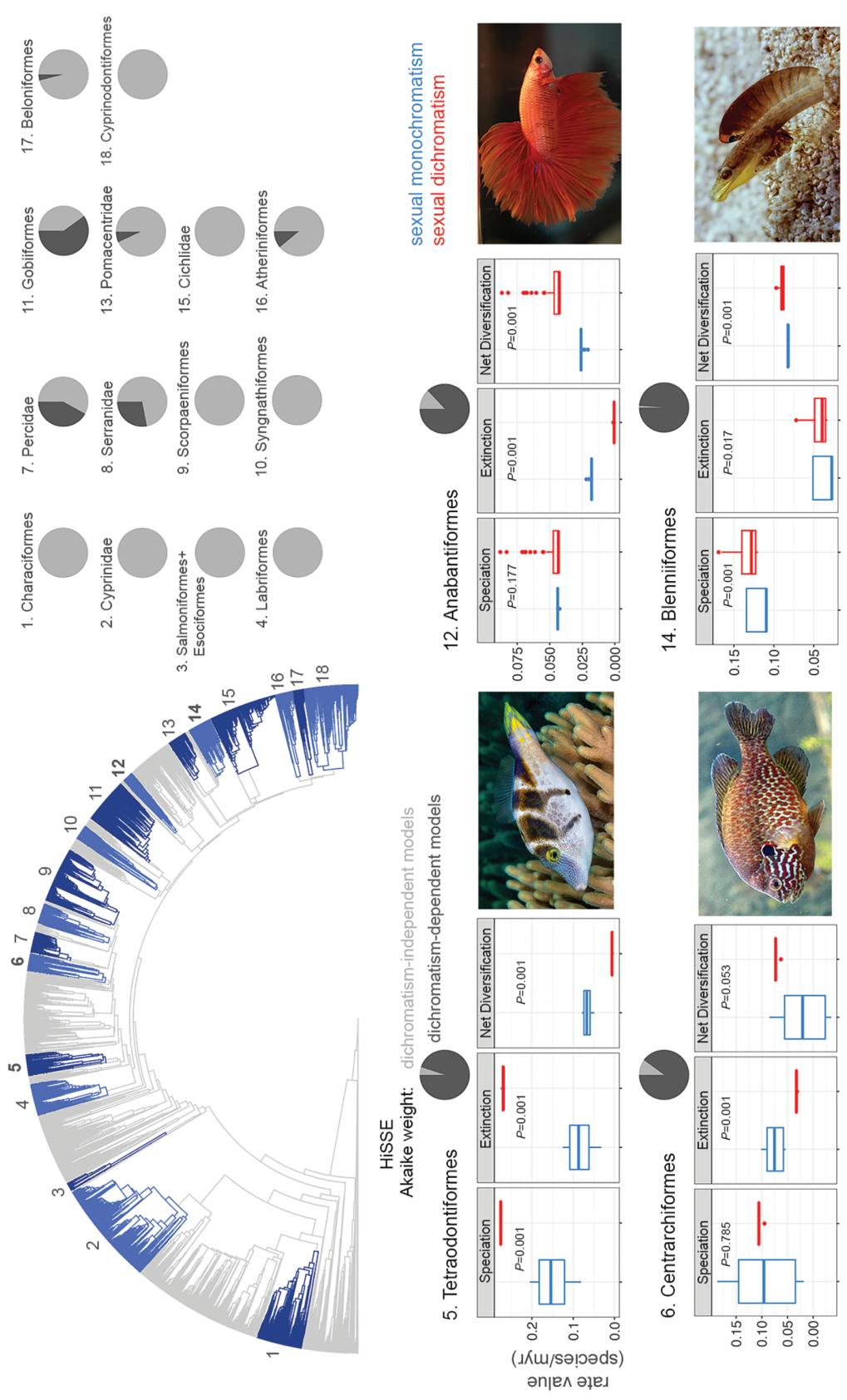


Figure 2: Sexual dichromatism and diversification within 18 fish clades. Clades chosen for analyses (table A5) are highlighted on the phylogeny in alternating shades of blue for visibil-ity. Pie diagrams indicate the relative Akaike weights (AICw) for 14 dichromatism-dependent hidden state speciation and extinction models (dark gray) versus nine dichromatism-independent models (light gray). Dichromatism-dependent models of diversification received >80% of the AICw support in four of 18 clades (*bottom*). Boxplots show model-averaged rates for individual species, including medians, first and third quartiles, and outliers (outside the 95% confidence interval). Rates for monochromatic species are shown in boxplots on the left in blue; rates for dichromatic species are shown on the right in red. Model fits are given in table A14. Model-averaged rates for all clades are given in table A15 and figs. A2–A5. Photograph credits: Tetraodontiformes: François Libert; Centrarchiformes: Wikimedia user Cephas, CC BY-SA 3.0; Anabantiformes: Flickr user Daniella Vereeken, CC BY 2.0; Blenniiformes: Betty Wills (Atsme), Wikimedia Commons, CC-BY-SA 4.0.

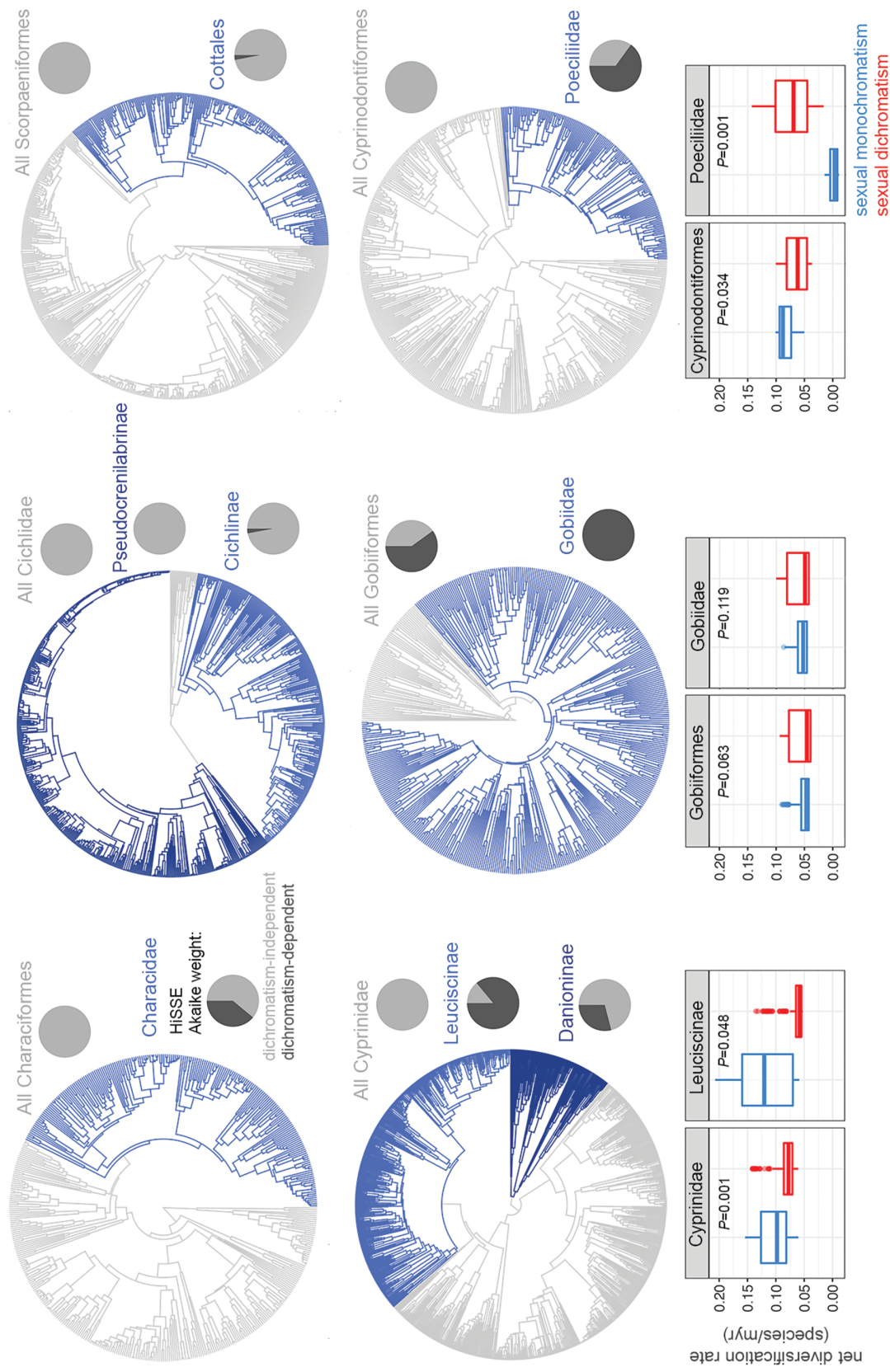


Figure 3: Sexual dichromatism and diversification at shallower phylogenetic scales (six subclades from the largest of the 18 clades in fig. 2). Pie diagrams indicate relative Akaike weights for 14 dichromatism-dependent hidden state speciation and extinction models (dark gray) versus nine dichromatism-independent models (light gray). The bottom row shows differences in rates between three subclades and their larger clades for which dichromatism-dependent models received the most support. Boxplots show model-averaged net diversification rates for individual species, including medians, first and third quartiles, and outliers (outside the 95% confidence interval). Rates for monochromatic species are shown in boxplots on the left in blue; rates for dichromatic species are shown on the right in red. Model fits are given in table A18. Model-averaged rates of speciation, extinction, and net diversification are given for all subclades in table A19 and fig. A6.

support for an effect of dichromatism, but there was similar support among the 40-, 60-, and 80-myrr subsets. There was no correlation between model support and the proportion of species sampled or the proportion of species with sexual dichromatism (table A28). Instead, model support was negatively correlated with clade size (fig. 4C). This pattern appeared when we pooled all 49 clades for comparison (clade size: Spearman's $\rho = -0.680$, $P < .001$; clade age: $\rho = -0.264$, $P = .073$) and also within the 100-, 80-, and 60-myrr subsets individually (table A28).

Among all rank-free clades, the largest clade with support for dichromatism-dependent diversification contained 1,107 described species (summed AICw = 72%). None of the 13 clades larger than this (1,442–4,139 species) had a summed AICw >5% for dichromatism-dependent models. Eleven clades had strong support for dichromatism-dependent models (AICw >80%). These clades ranged from 39.5 to 93.3 myrr old (mean = 62.5) and had 106–414 sampled tips (mean = 202.3) and 144–1,041 described species (mean = 556.6). Clades with equivocal support (AICw = 40%–70%) tended to be small (<200 tips, 167–339 described species). Putting these observations together, the power to detect dichromatism-dependent diversification was sufficient in clades with >300 species, yet it was unlikely to be detected in clades with >1,000 species.

We also considered whether the number of independent origins of monochromatism or dichromatism in a clade was related to the support for dichromatism-dependent models, as expected if the power to detect this effect increases with greater phylogenetic replication. For each clade, we counted the number of transitions to either state using ancestral-state reconstructions from the best-fitting HiSSE model and used the smaller of the two counts in correlations with model support. This relationship was negative, the opposite of our prediction ($\rho = -0.612$, $P < .001$; table A28). The number of origins was strongly correlated with the size of the phylogeny ($\rho = 0.804$, $P < .001$). These results further suggest that clade size is the most consistent predictor of support for dichromatism-dependent HiSSE models.

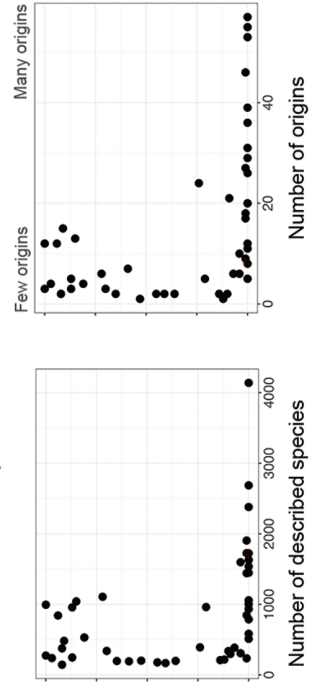
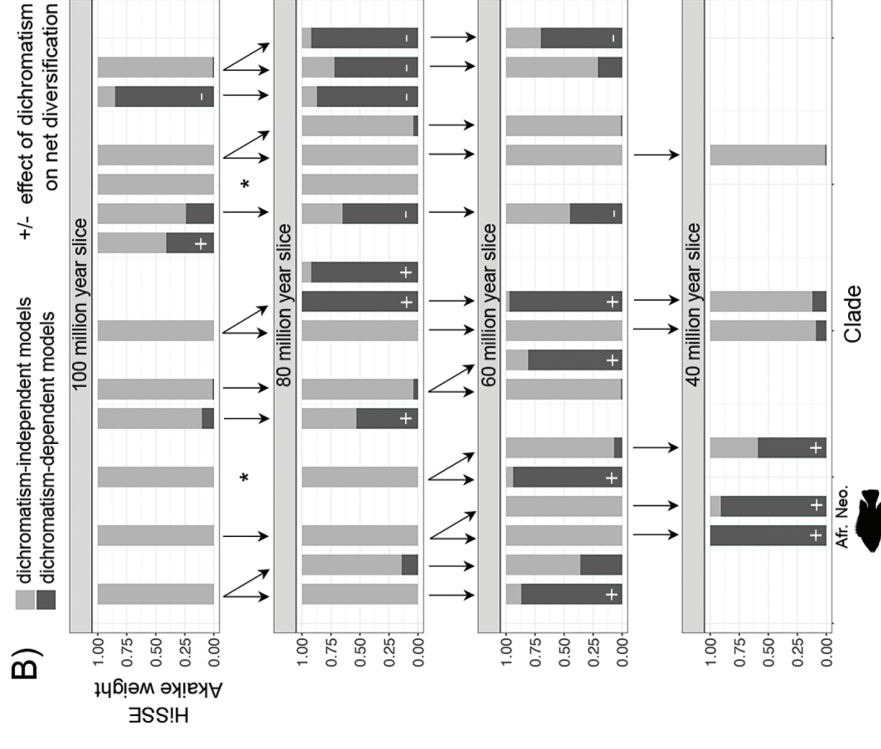
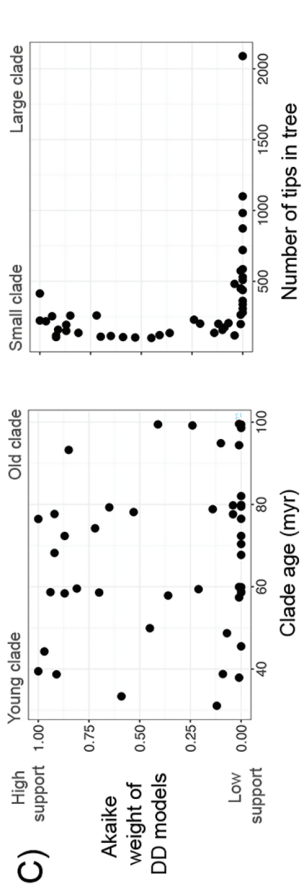
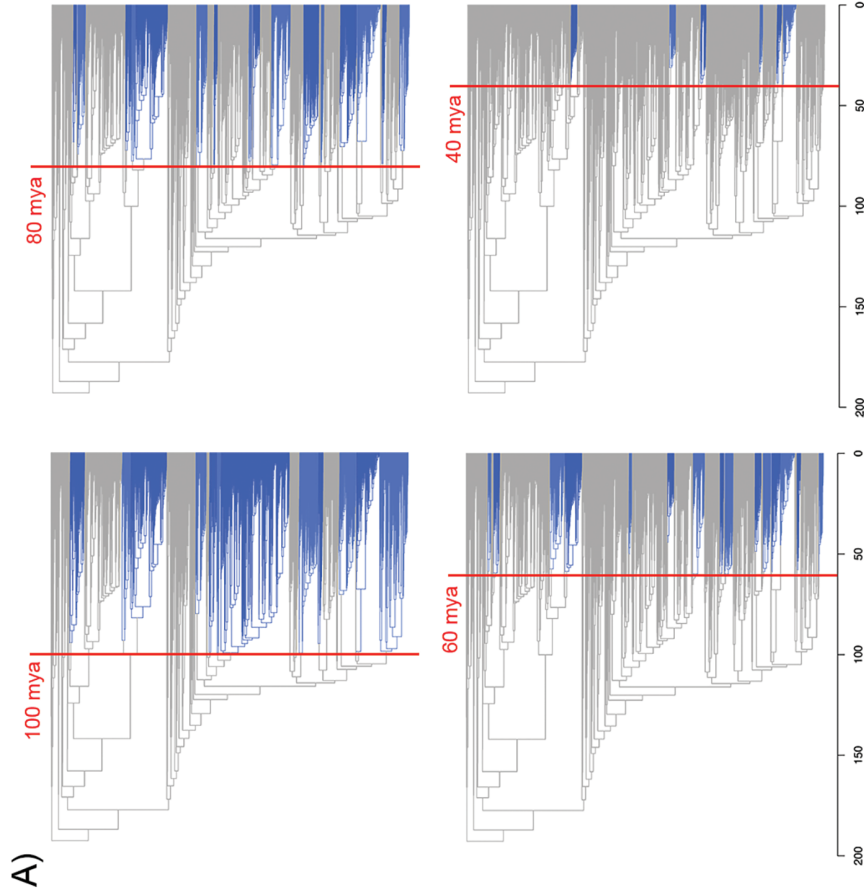
Discussion

A major goal in evolutionary biology is to identify the traits that influence diversification rate differences among clades (Wiens 2017). Since Darwin (1871), sexual dichromatism has been thought to be related to speciation and thereby to faster rates of net diversification (speciation minus extinction) and greater species richness (Lande 1981; West-Eberhard 1983; Ritchie 2007; Kraaijeveld et al. 2011; Boughman 2014). Classic studies in several fish families have provided strong evidence that sexual dichromatism is related to assortative mating (Houde and Endler 1990; Seehausen

et al. 1997; Elmer et al. 2009), reproductive isolation (Boughman 2001; Mendelson 2003; Seehausen et al. 2008), and diversification (Kazancıoğlu et al. 2009; Wagner et al. 2012; Smith et al. 2015; Furness et al. 2019; McGee et al. 2020). We assembled and analyzed a data set of unprecedented size to address how dichromatism impacts diversification rates across ray-finned fishes. Large data sets can reduce potential biases caused when well-studied groups are not broadly representative of all taxa (Beaulieu and O'Meara 2018). In addition, large data sets allowed us to test for scale dependency in the effects of dichromatism on diversification rates. Researchers now recognize that the factors that drive diversification may differ depending on the temporal or taxonomic scale examined (Jablonski and Bottjer 1991; Benton 2009; Kraaijeveld et al. 2011; Bellwood et al. 2017; Graham et al. 2018). Yet empirical demonstrations of scale-dependent effects of traits on diversification are rare (but see Huang and Rabosky 2014; Harrington and Reeder 2017; Hernández-Hernández and Wiens 2020).

Our results suggest that sexual dichromatism can explain diversification rate differences but only over limited phylogenetic scales. For example, we found evidence that dichromatism increases net diversification in the family Poeciliidae (fig. 3). However, we found no such relationship in the order Cyprinodontiformes, which contains Poeciliidae (fig. 3). Similarly, it is well established that nuptial colors help to maintain reproductive isolation among closely related cichlid species (Seehausen et al. 1997, 2008; Elmer et al. 2009), and sexual dichromatism can help explain which cichlid lineages have radiated in African lakes (Wagner et al. 2012; McGee et al. 2020). However, we found no support for dichromatism-dependent HiSSE models for Cichlidae or for its two major subfamilies (fig. 3). We eventually found strong support for a positive relationship between dichromatism and diversification in clades within each subfamily that were selected using a rank-free time cut-off of 40 million years (fig. 4B).

The scale-dependent effect of sexual dichromatism on diversification rates has been previously suggested. Kraaijeveld et al. (2011) compared different studies of birds and suggested that sexual dichromatism has the strongest effect at shallow scales (e.g., within species or genera; but see Cooney et al. 2017). They attributed this pattern to the extinction of newly formed species over time, eroding the signal of diversification driven by sexual selection. This mechanism should homogenize diversification rates and reduce rate disparities among older clades. However, our results suggest an additional cause for the scale-dependent effect (even if extinction is present). Specifically, the disparity in rates among fish clades was often greater than diversification rate variation within clades (e.g., rates among the 18 clades in figs. 2, A2). For example, even monochromatic cichlids have higher diversification rates than dichromatic



anabantiforms (fig. A2). Our rank-free comparisons also shed light on the cause of the scale-dependent effect. Support for dichromatism-dependent HiSSE models was negatively correlated with clade size, regardless of clade age (fig. 4C; table A27). The support for dichromatism-independent models for larger clades (figs. 2–4) indicates that other factors (hidden states) better explain diversification rate variation among subclades.

On the basis of these observations, we suggest that large clades are likely to be heterogeneous in other variables that drive diversification, and these variables could have stronger influence on diversification rates than sexual dichromatism. For example, habitat and trophic divergence are thought to precede divergence in sexual signals in vertebrate radiations (Mesnick 1996; Streebman and Danley 2003; Bellwood et al. 2017). These ecological factors may drive diversification rate differences among distantly related clades. Sexual dichromatism may then act on these background rates within clades. This top-down explanation is distinct from the bottom-up explanation (erosion of signal through time) of past studies (e.g., Kraaijeveld et al. 2011).

Many factors influence diversification rates across a large clade like actinopterygian fishes. Some readers may be concerned that no signal of any single trait on diversification rates could appear at this large scale because of confounding factors influencing diversification. However, several recent studies have shown significant effects of individual traits on speciation or net diversification rates across distantly related actinopterygian clades using similar data and methods as in this study. These traits include latitude (using HiSSE: Miller et al. 2018; using BAMM: Rabosky et al. 2018), diet (Siqueira et al. 2020), lacustrine habitat (Miller 2021), diadromy (Corush 2019), and several other variables (Tedesco et al. 2017). In addition, single traits (e.g., microhabitat) have been shown to have significant explanatory power at much deeper scales, including across all vertebrates (Wiens 2015) and all animals overall (Jezkova and Wiens 2017).

Why might some traits matter more than others over deep time? It may be that traits must have a consistent and replicated effect among clades for this effect to appear in

whole-clade analyses. For example, not only are high latitudes positively related to diversification rates in marine fishes, but also this positive effect appears independently in distantly related clades found near both the northern and the southern poles (Rabosky et al. 2018). In contrast, sexual dichromatism does not have an effect on diversification in all clades where it is present (even among young clades; figs. 3, 4). When it does, the effect can be positive or negative (figs. 2–4). Thus, there is no apparent effect when these clades are combined (fig. 1; table A4).

Our results have methodological implications for diversification rate studies. Analyses of very large clades are becoming increasingly common (e.g., Huang and Rabosky 2014; Miller et al. 2018; Rabosky et al. 2018; Hernández-Hernández and Wiens 2020). Greater sampling affords greater power to detect trait-dependent diversification (Davis et al. 2013) and uncover generalities beyond model clades (Beaulieu and O’Meara 2018). However, the strongest support for an effect of sexual dichromatism was found among clades with only ~200 tips (fig. 4C). This conflict may be relevant to other trait-dependent diversification studies. Counterintuitively, simply performing analyses on the largest possible phylogeny may improve power but lead to an incorrect conclusion that the trait is entirely irrelevant to diversification. We are not aware of existing comparative methods that can automatically detect the scale at which a trait is important for diversification. Therefore, we encourage researchers studying large clades to also perform analyses on a range of subclades.

Our results suggest that the presence of sexual dichromatism may not be enough to influence diversification rates by itself. There are several potential reasons for this. First, sexual dichromatism may promote speciation only when certain extrinsic and intrinsic conditions are met, such as environments with high water clarity (Seehausen et al. 1997, 2008; Hodge et al. 2020) or when male courtship or ornaments are also present (Furness et al. 2019). Second, some authors have suggested that the rate of change in sexual signals is what drives diversification (Gomes et al. 2016). Given this idea, we may not expect higher diversification rates in groups with sexual signals that

Figure 4: Sexual dichromatism and diversification with time. *A*, Summary of our rank-free clade selection procedure. We sliced the ray-finned fish phylogeny at 100, 80, 60, and 40 myr ago and collected the resulting subclades. Subclades that were suitable for hidden state speciation and extinction (HiSSE) analyses are highlighted in blue. Clade identities are given in tables A7–A10. *B*, Results of HiSSE analyses on rank-free clades. Bars show the relative Akaike weights for 14 dichromatism-dependent models (dark gray) versus nine dichromatism-independent models (light gray). Each bar represents one clade in *A*. Signs indicate whether sexual dichromatism has a positive (+) or negative (–) effect on net diversification. Full HiSSE results are given in tables A20–A27. Clades are positioned vertically such that nested subclades are below their parent clade, indicated with arrows. Some clades have more than one nested subclade (double arrows). An asterisk indicates that an identical clade was produced with the 80-myr slice as with the 100-myr slice because of the stem branch crossing both time slices. Fish icon denotes African and Neotropical cichlid clades (image from Phylopic, with credit to Milton Tan). *C*, Comparing the support for dichromatism-dependent HiSSE models when all 47 unique clades are pooled. We found a hook-shaped relationship with support and clade size. We also found this relationship using the number of origins (smaller count of the origins of monochromatism or dichromatism). Full correlation results are given in table A28.

are present but relatively invariant among species. Third, diversification rates may be associated with only elaborate dichromatism, if the degree of dimorphism is a proxy for the strength of sexual selection (Darwin 1871). We did not find support for this hypothesis in wrasses or darters (text A4; table A12; fig. A1). However, we have not tested this hypothesis among all fish clades. The three scenarios listed above may explain why diversification rates vary among dichromatic species. However, these factors cannot explain why monochromatic species often had higher diversification rates than dichromatic species.

Why might sexually monochromatic fishes have diversification rates that are similar to (or faster) than those of dichromatic fishes? In some clades, the absence of sexual dichromatism may not indicate the absence of assortative mating or sexual selection. Monochromatic fishes may have other features involved in species recognition or mate choice. Hamlets (*Serranidae*) have the fastest diversification rates among reef fishes (Siqueira et al. 2020). Sympatric species of hamlets are reproductively isolated by species-specific but sexually monochromatic color patterns (Hench et al. 2019). Butterflyfishes are another sexually monochromatic clade in which species-specific patterns may maintain species boundaries (Hemingson et al. 2019). Diversification rates among shallow marine fishes might be better explained by color in general rather than sexually dimorphic colors (Mesnick 1996; Bellwood et al. 2017). In environments with poor color transmission, signals that are bioluminescent, acoustic, tactile, and electrical are often involved in courtship and may also increase diversification rates (Mesnick 1996; Davis et al. 2014). In fact, these signals could have an even stronger and more consistent effect on diversification than color. Mesnick (1996) found that nonvisual sexual signals were associated with faster diversification in all fish clades with these signals. In contrast, visual signals had positive, negative, or no effect on diversification. The possibility remains that sexual selection in general will help explain variation in diversification rates among ray-finned fishes but only when all sexually selected signals are considered (Mesnick 1996).

Observations within fishes (e.g., assortative mating among populations) imply that dichromatism increases species richness by increasing speciation rates, as predicted by classic literature (Darwin 1871; Lande 1981; West-Eberhard 1983). Yet higher diversification rates in dichromatic lineages were due to lower extinction rates (not faster speciation rates) in several clades, including *Anabantiformes*, *Centrarchiformes*, and *Poeciliidae* (fig. 2; tables A14, A18). In some clades, dichromatism also decreased speciation (e.g., *Leuciscinae*) and increased extinction (e.g., *Blenniiformes*, *Tetraodontiformes*). There are several mechanisms that might explain these disparate effects. Sexual selection can increase extinction risk if sexually selected

traits increase mortality (Promislow et al. 1992) but can also reduce extinction by facilitating the purging of harmful mutations (Lumley et al. 2015). The effect of sexual selection on extinction in individual clades could depend on factors such as fecundity, population size, environmental stochasticity, and predation pressure (Promislow et al. 1992; Martínez-Ruiz and Knell 2017). Incorporating approaches from community ecology with diversification rate analyses may reveal the biological mechanisms for how sexual dichromatism influences speciation or extinction within clades (Tsuji and Fukami 2020).

Finally, what do the fish clades in which sexual dichromatism affects diversification have in common that is not shared by similarly sized clades in which dichromatism has no effect? Summarizing among all analyses (including ranked and rank-free clades), we found 13 independent clades with support for dichromatism-dependent HiSSE models (with either positive or negative effects) once the appropriate phylogenetic scale was determined. These clades were *Anabantiformes*, *Blenniiformes*, *Centrarchiformes*, two clades in *Characiformes*, two clades in *Cichlidae*, *Gobiidae*, *Leuciscinae*, *Percidae*, *Poeciliidae*, *Tetraodontiformes*, and a clade containing loaches. There were eight clades where dichromatism-dependent models were not supported even at smaller scales. These clades were *Atheriniformes*, *Beleniformes*, *Labridae*, *Pomacentridae*, *Salmoniformes* + *Esoiciformes*, *Scorpaeniformes*, *Serranidae*, and *Syngnathiformes*. Informal comparisons suggest two general differences between these groups (though there are exceptions). The first set of clades (with dichromatism-dependent diversification) are mostly freshwater, whereas the second set (dichromatism independent) are mostly marine. In addition, the first set of clades tend to show parental care (usually by males), whereas the second set generally lack parental care (Breder and Rosen 1966; Thresher 1984; Winemiller 1992). The codistribution of sexual dichromatism and parental care (Mank et al. 2005) could indicate stronger sexual selection than dichromatism alone if parental care is also under sexual selection (Alonzo 2012). Parental care may evolve in freshwater more often than in the ocean if pelagic eggs are adaptive in marine environments (Winemiller 1992). Further, gene flow from highly dispersive larvae may offset deme formation due to sexual selection on divergent male colors in marine species. Our data set on the presence of sexual dichromatism among fishes could be used to test these hypotheses in future comparative studies.

Conclusions

Researchers have found two patterns that potentially link sexual selection and speciation (Boughman 2014). The first is that premating isolation among many populations and

species is based on differences in secondary sexual characteristics, such as nuptial coloration (Seehausen et al. 1997; Boughman 2001; Mendelson 2003; Elmer et al. 2009). The second pattern is a positive relationship between sexual dimorphism and diversification rates detected using comparative methods (Barracough et al. 1995; Mesnick 1996; Mank 2007; Wagner et al. 2012; Furness et al. 2019). Our study suggests that the first pattern may not always lead to the second. On the basis of our results, the well-demonstrated effects of sexual dichromatism on speciation do not scale up to explain diversification rate variation across major clades (e.g., when comparing rates among monomorphic and dimorphic families and orders). Thus, our study helps resolve the paradox of the strong support for the effect of sexual selection on speciation from experimental and theoretical studies and the mixed support for this effect from macroevolutionary studies (Kraaijeveld et al. 2011; Tsuji and Fukami 2020).

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Statement of Authorship

E.C.M. conceptualized the work, led the data collection, designed and performed the analyses, and wrote the original manuscript. S.L.M. contributed to data collection. All authors helped to interpret the results and revise the manuscript.

Data and Code Availability

The complete dichromatism data set is included as data set S1. That data set as well as scripts needed to replicate analyses are deposited in the Dryad Digital Repository (<https://doi.org/10.5061/dryad.hqbzkh1bs>; Miller et al. 2021).

Literature Cited

Albert, A. Y. K., S. Sawaya, T. H. Vines, A. K. Knecht, C. T. Miller, B. R. Summers, S. Balabhadra, D. M. Kingsley, and D. Schluter.

2008. The genetics of adaptive shape shift in stickleback: pleiotropy and effect size. *Evolution* 62:76–85.
- Alonzo, S. H. 2012. Sexual selection favours male parental care, when females can choose. *Proceedings of the Royal Society B* 279:1784–1790.
- Amundsen, T. 2003. Fishes as models in studies of sexual selection and parental care. *Journal of Fish Biology* 63:17–52.
- Barracough, T. G., P. H. Harvey, and S. Nee. 1995. Sexual selection and taxonomic diversity in passerine birds. *Proceedings of the Royal Society B* 259:211–215.
- Beaulieu, J. M., and B. C. O'Meara. 2016. Detecting hidden diversification shifts in models of trait-dependent speciation and extinction. *Systematic Biology* 65:583–601.
- . 2018. Can we build it? yes we can, but should we use it? assessing the quality and value of a very large phylogeny of campanulid angiosperms. *American Journal of Botany* 105:417–432.
- Bellwood, D. R., C. H. R. Goatley, and O. Bellwood. 2017. The evolution of fishes and corals on reefs: form, function and interdependence. *Biological Reviews* 92:878–901.
- Benton, M. J. 2009. The Red Queen and the Court Jester: species diversity and the role of biotic and abiotic factors through time. *Science* 323:728–732.
- Boettiger, C., D. T. Lang, and P. C. Wainwright. 2012. rfishbase: exploring, manipulating and visualizing FishBase data from R. *Journal of Fish Biology* 81:2030–2039.
- Bolnick, D. I., and M. Doebeli. 2003. Sexual dimorphism and adaptive speciation: two sides of the same ecological coin. *Evolution* 57:2433–2449.
- Bossu, C. M., and T. J. Near. 2015. Ecological constraint and the evolution of sexual dichromatism in darters. *Evolution* 69:1219–1231.
- Boughman, J. W. 2001. Divergent sexual selection enhances reproductive isolation in sticklebacks. *Nature* 411:944–948.
- . 2014. Speciation by sexual selection. Pages 520–528 *in* J. B. Losos, ed. *Princeton guide to evolution*. Princeton University Press, Princeton, NJ.
- Breder, C. M., and D. E. Rosen. 1966. *Modes of reproduction in fishes*. TFH, Neptune City, NJ.
- Burnham, K. P., and D. R. Anderson. 2002. *Model selection and multimodel inference: a practical information-theoretic approach*. Springer, New York.
- Burriss, E. D., and M. Tan. 2017. Ecological opportunity alters the timing and shape of adaptive radiation. *Evolution* 71:2650–2660.
- Caetano, D. S., B. C. O'Meara, and J. M. Beaulieu. 2018. Hidden state models improve state-dependent diversification approaches, including biogeographical models. *Evolution* 72:2308–2324.
- Choat, J. H., O. S. Klanten, L. V. Herwerden, D. R. Robertson, and K. D. Clements. 2012. Patterns and processes in the evolutionary history of parrotfishes (family Labridae). *Biological Journal of the Linnean Society* 107:529–557.
- Cooney, C. R., J. A. Tobias, J. T. Weir, C. A. Botero, and N. Seddon. 2017. Sexual selection, speciation and constraints on geographical range overlap in birds. *Ecology Letters* 20:863–871.
- Corush, J. B. 2019. Evolutionary patterns of diadromy in fishes: more than a transitional state between marine and freshwater. *BMC Evolutionary Biology* 19:168.
- Darwin, C. 1871. *The descent of man and selection in relation to sex*. J. Murray, London.
- Davis, M. P., N. I. Holcroft, E. O. Wiley, J. S. Sparks, and W. Leo Smith. 2014. Species-specific bioluminescence facilitates speciation in the deep sea. *Marine Biology* 161:1139–1148.

- Davis, M. P., P. E. Midford, and W. Maddison. 2013. Exploring power and parameter estimation of the BiSSE method for analyzing species diversification. *BMC Evolutionary Biology* 13:38.
- Eaton, M. D. 2005. Human vision fails to distinguish widespread sexual dichromatism among sexually “monochromatic” birds. *Proceedings of the National Academy of Sciences of the USA* 102:10942–10946.
- Elmer, K. R., T. K. Lehtonen, and A. Meyer. 2009. Color assortative mating contributes to sympatric divergence of Neotropical cichlid fish. *Evolution* 63:2750–2757.
- Freckleton, R. P., P. H. Harvey, and M. Pagel. 2002. Phylogenetic analysis and comparative data: a test and review of evidence. *American Naturalist* 160:712–726.
- Froese, R., and D. Pauly. 2019. FishBase. Version 08/2019. <http://www.fishbase.org/>.
- Furness, A. I., B. J. A. Pollux, R. W. Meredith, M. S. Springer, and D. N. Reznick. 2019. How conflict shapes evolution in poeciliid fishes. *Nature Communications* 10:3335.
- Garland, T., A. W. Dickerman, C. M. Janis, and J. A. Jones. 1993. Phylogenetic analysis of covariance by computer simulation. *Systematic Biology* 42:265–292.
- Gomes, A. C. R., M. D. Sorenson, and G. C. Cardoso. 2016. Speciation is associated with changing ornamentation rather than stronger sexual selection. *Evolution* 70:2823–2838.
- Graham, C. H., D. Storch, and A. Machac. 2018. Phylogenetic scale in ecology and evolution. *Global Ecology and Biogeography* 27:175–187.
- Harrington, S., and T. W. Reeder. 2017. Rate heterogeneity across Squamata, misleading ancestral state reconstruction and the importance of proper null model specification. *Journal of Evolutionary Biology* 30:313–325.
- Helfman, G. S., B. B. Collette, D. E. Facey, and B. W. Bowen. 2009. *The diversity of fishes: biology, evolution, and ecology*. 2nd ed. Wiley-Blackwell, Hoboken, NJ.
- Hemingson, C. R., P. F. Cowman, J. R. Hodge, and D. R. Bellwood. 2019. Colour pattern divergence in reef fish species is rapid and driven by both range overlap and symmetry. *Ecology Letters* 22:190–199.
- Hench, K., M. Vargas, M. P. Höppner, W. O. McMillan, and O. Puebla. 2019. Inter-chromosomal coupling between vision and pigmentation genes during genomic divergence. *Nature Ecology and Evolution* 3:657–667.
- Hernández-Hernández, T., and J. J. Wiens. 2020. Why are there so many flowering plants? a multiscale analysis of plant diversification. *American Naturalist* 195:948–963.
- Hodge, J. R., F. Santini, and P. C. Wainwright. 2020. Colour dimorphism in labrid fishes as an adaptation to life on coral reefs. *Proceedings of the Royal Society B* 287:20200167.
- Houde, A. E., and J. A. Endler. 1990. Correlated evolution of female mating preferences and male color patterns in the guppy *Poecilia reticulata*. *Science* 248:1405–1408.
- Huang, H., and D. L. Rabosky. 2014. Sexual selection and diversification: reexamining the correlation between dichromatism and speciation rate in birds. *American Naturalist* 184:E101–E114.
- Irwin, D. E. 2020. Assortative mating in hybrid zones is remarkably ineffective in promoting speciation. *American Naturalist* 195: E150–E167.
- Jablonski, D., and D. J. Bottjer. 1991. Environmental patterns in the origins of higher taxa: the post-Paleozoic fossil record. *Science* 252:1831–1833.
- Jezkova, T., and J. J. Wiens. 2017. What explains patterns of diversification and richness among animal phyla? *American Naturalist* 189:201–212.
- Kazancıoğlu, E., T. J. Near, R. Hanel, and P. C. Wainwright. 2009. Influence of sexual selection and feeding functional morphology on diversification rate of parrotfishes (Scaridae). *Proceedings of the Royal Society B* 276:3439–3446.
- Kodric-Brown, A. 1990. Mechanisms of sexual selection: insights from fishes. *Annales Zoologici Fennici* 27:87–100.
- . 1998. Sexual dichromatism and temporary color changes in the reproduction of fishes. *Integrative and Comparative Biology* 38:70–81.
- Kraaijeveld, K., F. J. L. Kraaijeveld-Smit, and M. E. Maan. 2011. Sexual selection and speciation: the comparative evidence revisited. *Biological Reviews* 86:367–377.
- Lande, R. 1981. Models of speciation by sexual selection on polygenic traits. *Proceedings of the National Academy of Sciences of the USA* 78:3721–3725.
- Lorch, P. D., S. Proulx, L. Rowe, and T. Day. 2003. Condition-dependent sexual selection can accelerate adaptation. *Evolutionary Ecology Research* 5:867–881.
- Losey, G. S., T. W. Cronin, T. H. Goldsmith, D. Hyde, N. J. Marshall, and W. N. McFarland. 1999. The UV visual world of fishes: a review. *Journal of Fish Biology* 54:921–943.
- Louca, S., and M. W. Pennell. 2020. Extant timetrees are consistent with a myriad of diversification histories. *Nature* 580:502–505.
- Lumley, A. J., Ł. Michalczyk, J. J. N. Kitson, L. G. Spurgin, C. A. Morrison, J. L. Godwin, M. E. Dickinson, et al. 2015. Sexual selection protects against extinction. *Nature* 522:470–473.
- Mank, J. E. 2007. Mating preferences, sexual selection and patterns of cladogenesis in ray-finned fishes. *Journal of Evolutionary Biology* 20:597–602.
- Mank, J. E., D. E. L. Promislow, and J. C. Avise. 2005. Phylogenetic perspectives in the evolution of parental care in ray-finned fishes. *Evolution* 59:1570–1578.
- Martin, M. D., and T. C. Mendelson. 2014. Changes in sexual signals are greater than changes in ecological traits in a dichromatic group of fishes. *Evolution* 68:3618–3628.
- Martínez-Ruiz, C., and R. J. Knell. 2017. Sexual selection can both increase and decrease extinction probability: reconciling demographic and evolutionary factors. *Journal of Animal Ecology* 86:117–127.
- Martins, M. J. F., T. M. Puckett, R. Lockwood, J. P. Swaddle, and G. Hunt. 2018. High male sexual investment as a driver of extinction in fossil ostracods. *Nature* 556:366–369.
- Matzke, N. J. 2014. Model selection in historical biogeography reveals that founder-event speciation is a crucial process in island clades. *Systematic Biology* 63:951–970.
- McGee, M. D., S. R. Borstein, J. I. Meier, D. A. Marques, S. Mwaiko, A. Taabu, M. A. Kishe, et al. 2020. The ecological and genomic basis of explosive adaptive radiation. *Nature* 586:75–79.
- Mendelson, T. C. 2003. Sexual isolation evolves faster than hybrid inviability in a diverse and sexually dimorphic genus of fish (Percidae: *Etheostoma*). *Evolution* 57:317–327.
- Mesnick, S. L. 1996. *Sexual selection and biological diversification: patterns and processes*. PhD diss. University of Arizona, Tucson.
- Meyer, A. L. S., and J. J. Wiens. 2018. Estimating diversification rates for higher taxa: BAMM can give problematic estimates of rates and rate shifts. *Evolution* 72:39–53.

- Miller, E. C. 2021. Comparing diversification rates in lakes, rivers, and the sea. *bioRxiv*, <https://doi.org/10.1101/2020.05.24.113761>.
- Miller, E. C., K. T. Hayashi, D. Song, and J. J. Wiens. 2018. Explaining the ocean's richest biodiversity hotspot and global patterns of fish diversity. *Proceedings of the Royal Society B* 285:20181314.
- Miller, E. C., S. L. Mesnick, and J. J. Wiens. 2021. Data from: Sexual dichromatism is decoupled from diversification over deep time in fishes. *American Naturalist*, Dryad Digital Repository, <https://doi.org/10.5061/dryad.hqzbkxh1bs>.
- Moore, B. R., S. Höhna, M. R. May, B. Rannala, and J. P. Huelsenbeck. 2016. Critically evaluating the theory and performance of Bayesian analysis of macroevolutionary mixtures. *Proceedings of the National Academy of Sciences of the USA* 113:9569–9574.
- Morrow, E. H., T. E. Pitcher, and G. Arnqvist. 2003. No evidence that sexual selection is an “engine of speciation” in birds. *Ecology Letters* 6:228–234.
- Poe, S. C., Anderson, and J. Barnett. 2020. On the selection and analysis of clades in comparative evolutionary studies. *Systematic Biology* 70:190–196.
- Promislow, D. E. L., R. Montgomerie, and T. E. Martin. 1992. Mortality costs of sexual dimorphism in birds. *Proceedings of the Royal Society B* 250:143–150.
- Rabosky, D. L. 2014. Automatic detection of key innovations, rate shifts, and diversity-dependence on phylogenetic trees. *PLoS ONE* 9:e89543.
- . 2015. No substitute for real data: a cautionary note on the use of phylogenies from birth-death polytomy resolvers for downstream comparative analyses. *Evolution* 69:3207–3216.
- Rabosky, D. L., J. Chang, P. O. Title, P. F. Cowman, L. Sallan, M. Friedman, K. Kaschner, et al. 2018. An inverse latitudinal gradient in speciation rate for marine fishes. *Nature* 559:392–395.
- Rabosky, D. L., and E. E. Goldberg. 2015. Model inadequacy and mistaken inferences of trait-dependent speciation. *Systematic Biology* 64:340–355.
- Rabosky, D. L., M. Grundler, C. Anderson, P. Title, J. J. Shi, J. W. Brown, H. Huang, and J. G. Larson. 2014. BAMMtools: an R package for the analysis of evolutionary dynamics on phylogenetic trees. *Methods in Ecology and Evolution* 5:701–707.
- Rabosky, D. L., and H. Huang. 2016. A robust semi-parametric test for detecting trait-dependent diversification. *Systematic Biology* 65:181–193.
- Rabosky, D. L., J. S. Mitchell, and J. Chang. 2017. Is BAMM flawed? theoretical and practical concerns in the analysis of multi-rate diversification models. *Systematic Biology* 66:477–498.
- Revell, L. J. 2012. *phytools: an R package for phylogenetic comparative biology (and other things)*. *Methods in Ecology and Evolution* 3:217–223.
- Reznick, D. N., A. I. Furness, R. W. Meredith, and M. S. Springer. 2017. The origin and biogeographic diversification of fishes in the family Poeciliidae. *PLoS ONE* 12:e0172546.
- Ritchie, M. G. 2007. Sexual selection and speciation. *Annual Review of Ecology, Evolution, and Systematics* 38:79–102.
- Schweikert, L. E., R. R. Fitak, E. M. Caves, T. T. Sutton, and S. Johnsen. 2018. Spectral sensitivity in ray-finned fishes: diversity, ecology and shared descent. *Journal of Experimental Biology* 221:jeb189761.
- Seehausen, O., Y. Terai, I. S. Magalhaes, K. L. Carleton, H. D. J. Mrosso, R. Miyagi, I. van der Sluijs, et al. 2008. Speciation through sensory drive in cichlid fish. *Nature* 455:620–626.
- Seehausen, O., J. van Alphen, and F. Witte. 1997. Cichlid fish diversity threatened by eutrophication that curbs sexual selection. *Science* 277:1808–1811.
- Servedio, M. R., and R. Burger. 2014. The counterintuitive role of sexual selection in species maintenance and speciation. *Proceedings of the National Academy of Sciences of the USA* 111:8113–8118.
- Siebeck, U. E., and N. J. Marshall. 2001. Ocular media transmission of coral reef fish—can coral reef fish see ultraviolet light? *Vision Research* 41:133–149.
- Siebeck, U. E., A. N. Parker, D. Sprenger, L. M. Mähger, and G. Wallis. 2010. A species of reef fish that uses ultraviolet patterns for covert face recognition. *Current Biology* 20:407–410.
- Siqueira, A. C., R. A. Morais, D. R. Bellwood, and P. F. Cowman. 2020. Trophic innovations fuel reef fish diversification. *Nature Communications* 11:2669.
- Siqueira, A. C., L. G. R. Oliveira-Santos, P. F. Cowman, and S. R. Floeter. 2016. Evolutionary processes underlying latitudinal differences in reef fish biodiversity. *Global Ecology and Biogeography* 25:1466–1476.
- Smith, A. J., N. Nelson-Maney, K. J. Parsons, W. J. Cooper, and R. C. Albertson. 2015. Body shape evolution in sunfishes: divergent paths to accelerated rates of speciation in the Centrarchidae. *Evolutionary Biology* 42:283–295.
- Streebman, T. J., and P. D. Danley. 2003. The stages of vertebrate evolutionary radiation. *Trends in Ecology and Evolution* 18:126–131.
- Tedesco, P. A., E. Paradis, C. Lévêque, and B. Hugué. 2017. Explaining global-scale diversification patterns in actinopterygian fishes. *Journal of Biogeography* 44:773–783.
- Thresher, R. E. 1984. *Reproduction in reef fishes*. TFH, Neptune City, NJ.
- Tsuji, K., and T. Fukami. 2020. Sexual dimorphism and species diversity: from clades to sites. *Trends in Ecology and Evolution* 35:105–114.
- Wagner, C. E., L. J. Harmon, and O. Seehausen. 2012. Ecological opportunity and sexual selection together predict adaptive radiation. *Nature* 487:366–369.
- West-Eberhard, M. J. 1983. Sexual selection, social competition, and speciation. *Quarterly Review of Biology* 58:155–183.
- Wiens, J. J. 2015. Explaining large-scale patterns of vertebrate diversity. *Biology Letters* 11:20150506.
- . 2017. What explains patterns of biodiversity across the Tree of Life? *BioEssays* 39:1600128.
- Winemiller, K. O. 1992. Life-history strategies and the effectiveness of sexual selection. *Oikos* 63:318–327.

References Cited Only in the Online Enhancements

- Abel, E. F. 1993. Colouration phenomena of Mediterranean blennies (Pisces, Blenniidae). *Marine Ecology* 14:291–312.
- Allen, G. R. 2009. *Field guide to marine fishes of tropical Australia and South-East Asia*. Western Australian Museum, Perth.
- Allen, G. R., and M. V. Erdmann. 2012. *Reef fishes of the East Indies*. 2nd ed. University of Hawaii Press, Honolulu.
- Allen, G. R., S. H. Midgley, and M. Allen. 2002. *Field guide to the freshwater fishes of Australia*. Western Australian Museum, Perth.
- Amcoff, M., A. Gonzalez-Voyer, and N. Kolm. 2013. Evolution of egg dummies in Tanganyikan cichlid fishes: the roles of parental care and sexual selection. *Journal of Evolutionary Biology* 26:2369–2382.
- Anderson, W. D., Jr. 1970. Revision of the genus *Symphysanodon* (Pisces: Lutjanidae) with descriptions of four new species. *Fishery Bulletin* 68:325–346.
- Andrews, C., and U. Baensch. 1991. *Tropical aquarium fish: comprehensive edition*. Tetra, Germany.

- Arai, H., and T. Sato. 2007. Prominent ornaments and rapid color change: use of horns as a social and reproductive signal in unicornfish (Acanthuridae: *Naso*). *Ichthyological Research* 54:49–54.
- Axelrod, H. R. 1993. The most complete colored lexicon of cichlids: every known cichlid illustrated in color. TFH, Neptune City, NJ.
- Ayling, T., and G. J. Cox. 1982. Collins guide to the sea fishes of New Zealand. William Collins, Auckland.
- Barbour, C. D., and R. R. Miller. 1978. A revision of the Mexican cyprinid fish genus *Algansea*. Miscellaneous publication no. 155, University of Michigan Museum of Zoology.
- Barlow, G. W. 1974. Contrasts in social behavior between Central American cichlid fishes and coral-reef surgeon fishes. *American Zoologist* 14:9–34.
- Bell-Cross, G., and Minshull, J. L. 1988. The fishes of Zimbabwe. Trustees of the National Museums and Monuments of Zimbabwe, Zimbabwe.
- Bianco, P. G., and V. Ketmaier. 2014. A revision of the *Rutilus* complex from Mediterranean Europe with description of a new genus, *Sarmarutilus*, and a new species, *Rutilus stoumboudae* (Teleostei: Cyprinidae). *Zootaxa* 3:379–402.
- Bills, I. R., and O. L. F. Weyl. 2002. A new species of the genus *Chetia* (Teleostei: Cichlidae) from the Lecitu River, Buzi System, Mozambique. *Smithiana Special Publication 1*, South African Institute for Aquatic Biodiversity.
- Borstein, S. Sam Borstein's cichlids. <http://www.borstein.info/>.
- Brantley, R. K., J. C. Wingfield, and A. H. Bass. 1993. Sex steroid levels in *Porichthys notatus*, a fish with alternative reproductive tactics, and a review of the hormonal bases for male dimorphism among teleost fishes. *Hormones and Behavior* 27:332–347.
- Bray, D. J., and M. F. Gomon, eds. 2019. Fishes of Australia. Museums Victoria and OzFishNet. <http://fishesofaustralia.net.au/>.
- Brichard, P. 1978. Fishes of Lake Tanganyika. TFH, Hong Kong.
- . 1989. Book of cichlids and all the other fishes of Lake Tanganyika. TFH, Neptune City, NJ.
- Brittan, M. R. 1972. *Rasbora*: a revision of the Indo-Malayan freshwater fish genus *Rasbora*. TFH, Hong Kong.
- Burgess, W. E. 1989. An atlas of freshwater and marine catfishes: a preliminary survey of the Siluriformes. TFH, Neptune City, NJ.
- Burgess, W. E., and H. R. Axelrod. 1973. Fishes of southern Japan and the Ryukyus. 1. Pacific marine fishes. TFH, Hong Kong.
- Butler, J. L., M. S. Love, and T. E. Laidig. 2012. A guide to the rockfishes, thornyheads, and scorpionfishes of the northeast Pacific. University of California Press, Los Angeles.
- Casazza, T. L., S. W. Ross, A. M. Necaie, and K. J. Sulak. 2005. Reproduction and mating behavior of the Atlantic flyingfish, *Cheilopogon melanurus* (Exocoetidae), off North Carolina. *Bulletin of Marine Science* 77:363–375.
- Chang, C.-H., F. Li, K.-T. Shao, Y.-S. Lin, T. Morosawa, S. Kim, H. Koo, et al. 2014. Phylogenetic relationships of Acheilognathidae (Cypriniformes: Cyprinoidea) as revealed from evidence of both nuclear and mitochondrial gene sequence variation: evidence for necessary taxonomic revision in the family and the identification of cryptic species. *Molecular Phylogenetics and Evolution* 81:182–194.
- Chen, I., J. Wu, and C. Hsu. 2008. The taxonomy and phylogeny of *Candidia* (Teleostei: Cyprinidae) from Taiwan, with description of a new species and comments on a new genus. *Raffles Bulletin of Zoology* 19:203–214.
- Cichlid-Forum. <https://www.cichlid-forum.com/>.
- Clemens, W. A., and G. V. Wilby. 1961. Fishes of the Pacific Coast of Canada. Vol. 68. 2nd ed. Fisheries Research Board of Canada, Ottawa.
- Coad, B. W. 2010. Freshwater fishes of Iraq. Pensoft, Sofia-Moscow.
- . 2014. Fishes of Afghanistan. Pensoft, Sofia-Moscow.
- Colin, P. L., and L. J. Bell. 1991. Aspects of the spawning of labrid and scarid fishes (Pisces: Labroidae) at Enewetak Atoll, Marshall Islands with notes on other families. *Environmental Biology of Fishes* 31:229–260.
- Collins, R. A., K. F. Armstrong, R. Meier, Y. Yi, S. D. J. Brown, R. H. Cruickshank, S. Keeling, and C. Johnston. 2012. Barcoding and border biosecurity: identifying cyprinid fishes in the aquarium trade. *PLoS ONE* 7:e28381.
- Conkel, D. 1993. Cichlids of North and Central America. TFH, Neptune City, NJ.
- Cooke, S. J., and D. P. Philipp. 2009. Centrarchid fishes: diversity, biology, and conservation. Wiley-Blackwell, Chichester.
- Cousseau, M. B., and R. G. Perrotta. 2000. Peces marinos de Argentina. Instituto Nacional de Investigación y Desarrollo Pesquero, Mar del Plata.
- Cowman, P. F., and D. R. Bellwood. 2011. Coral reefs as drivers of cladogenesis: expanding coral reefs, cryptic extinction events, and the development of biodiversity hotspots. *Journal of Evolutionary Biology* 24:2543–2562.
- Craig, M. T., Y. J. Sadovy de Mitcheson, and P. C. Heemstra. 2011. Groupers of the world: a field and market guide. NISC, Grahamstown.
- Cross, N. J., and G. R. Allen. 1982. Rainbowfishes of Australia and Papua New Guinea. TFH, Australia.
- Darshan, A., S. Abujam, and D. N. Das. 2018. Biodiversity of fishes in Arunachal Himalaya: systematics, classification, and taxonomic identification. Academic Press, London.
- DeMartini, E. 1985. Social behavior and coloration changes in painted greenling, *Oxylebias pictus* (Pisces: Hexagrammidae). *Copeia* 1985:966–975.
- De Pinna, M. C. C., C. J. Ferraris Jr., and R. P. Vari. 2007. A phylogenetic study of the Neotropical catfish family Cetopsidae (Osteichthyes, Ostariophysi, Siluriformes), with a new classification. *Zoological Journal of the Linnean Society* 150:755–813.
- Ebert, K. 2001. The puffers of fresh and brackish waters. Aqualog, Rodgau.
- Eccles, D. H., and E. Trewavas. 1989. Malawian cichlid fishes: the classification of some Haplochromine genera. Lake Fish Movies, Herten.
- Erismann, B. E., and L. G. Allen. 2005. Color patterns and associated behaviors in the kelp bass, *Paralabrax clathratus* (Teleostei: Serranidae). *Bulletin of the Southern California Academy of Sciences* 104:45–62.
- Eschmeyer, W. N., E. S. Herald, and H. Hammann. 1983. A field guide to Pacific Coast fishes of North America. Houghton Mifflin, New York.
- Ferraris, C. J., Jr., and R. P. Vari. 2007. Revision of catfishes of the genus *Eutropiichthys*, with the description of two new species (Siluriformes: Schilbidae). *Copeia* 2007:866–885.
- Fowler, H. W. 1972. A synopsis of the fishes of China. Antiquariaat Junk, Lochem.
- Fryer, G., and T. D. Iles. 1972. The cichlid fishes of the Great Lakes of Africa. TFH, Neptune City, NJ.
- Garcia, C. M. 2014. Mode of reproduction, mate choice, and species richness in Goodeid fish. Pages 253–288 in R. H. Macedo and

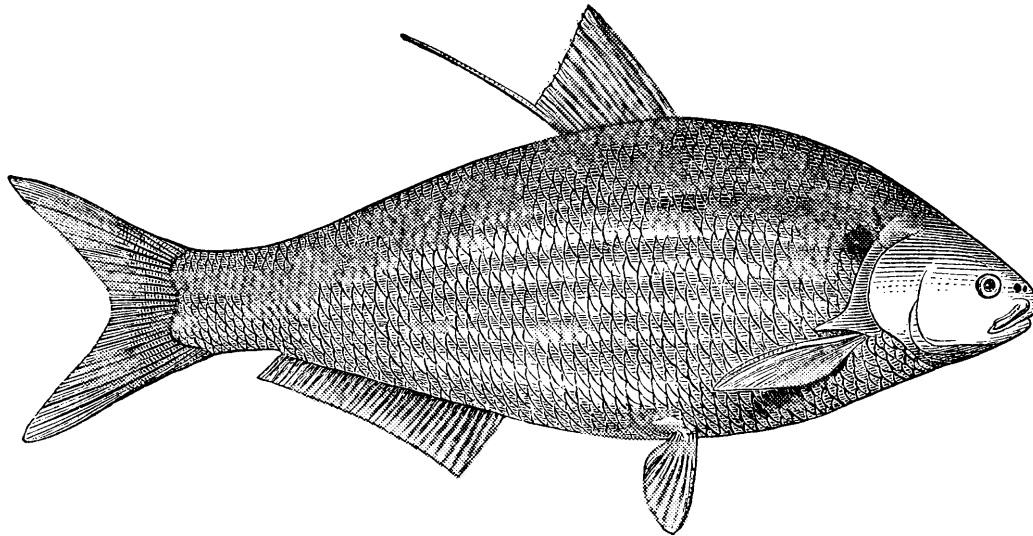
- G. Machado, eds. Sexual selection: perspectives and models from the Neotropics. Academic Press, Amsterdam.
- Goldstein, R. J. 2015. The betta handbook. Barron's Educational Series, New York.
- Gomon, M. F. 2006. A revision of the labrid fish genus *Bodianus* with descriptions of eight new species. Records of the Australian Museum Supplement 30:1–133.
- . 2017. A review of the tuskfishes, genus *Choerodon* (Labridae, Perciformes), with descriptions of three new species. Memoirs of Museum Victoria 76:1–111.
- Gonzalez-Voyer, A., J. L. Fitzpatrick, and N. Kolm. 2008. Sexual selection determines parental care patterns in cichlid fishes. Evolution 62:2015–2026.
- Hastings, P. A. 2019. Evolution of sexual dimorphism in tube blennies (Teleostei: Chaenopsidae). Integrative Organismal Biology 1:obz003.
- Hay, M. S., and I. J. Hodgkiss. 1981. Hong Kong freshwater fishes. Wishing, Hong Kong.
- Holleman, W. 2005. A review of the triplefin fish genus *Enneapterygius* (Blennioidei: Tripterygiidae) in the western Indian Ocean, with descriptions of four new species. Smithiana Bulletin 5:1–25.
- Huber, J., ed. Killi-data online. Accessed May 2019. <http://www.killi-data.org/>.
- Jones, A. G., and J. C. Avise. 1997. Polygynandry in the dusky pipefish *Syngnathus floridae* revealed by microsatellite DNA markers. Evolution 51:1611–1622.
- Kawase, H. 1998. Reproductive behavior and evolution of triggerfish (Balistidae) and filefish (Monacanthidae). Japanese Journal of Ichthyology 45:1–19.
- Kawase, S., and K. Hosoya. 2015. *Pseudorasbora pugnax*, a new species of minnow from Japan, and redescription of *P. pumila* (Teleostei: Cyprinidae). Ichthyological Exploration of Freshwaters 25:289–298.
- Kazancioglu, E., and S. H. Alonzo. 2010. A comparative analysis of sex change in Labridae supports the size advantage hypothesis. Evolution 64:2254–2264.
- King, D. 1996. Reef fishes and corals: east coast of southern Africa. Struik Nature, Cape Town.
- King, D., and V. Fraser. 2002. More reef fishes and nudibranchs: east and south coast of Southern Africa. Struik Nature, Cape Town.
- Kolpakov, E. V. 2011. Biology of Volk's sculpin *Cottus volki* (Cottidae) from the Serebryanka River (Central Primor'e). Journal of Ichthyology 51:178–186.
- Konings, A. 1990. Koning's book of cichlids and all the other fishes of Lake Malawi. TFH, Neptune City, NJ.
- Kortmulder, K. 1972. A comparative study in color patterns and behavior in seven Asiatic *Barbus* species (Cyprinidae, Ostariophysi, Osteichthys), a progress report. Behavior 19:1–331.
- Kottelat, M. 1990. Indochinese nemacheilines: a revision of nemacheiline loaches (Pisces: Cypriniformes) of Thailand, Burma, Laos, Cambodia and southern Viet Nam. Pfeil, Munchen.
- . 2001. Fishes of Laos. WHT, Colombo.
- Kottelat, M., and J. Freyhof. 2007. Handbook of European freshwater fishes. Publications Kottelat, Cornol.
- Kuiter, R. H. 1997. A photographic guide to the sea fishes of Australia. New Holland, Sydney.
- . 2000. Seahorses, pipefishes, and their relatives: a comprehensive guide to Syngnathiformes. TMC, Chorleywood.
- . 2002. Fairy and rainbow wrasses and their relatives: a comprehensive guide to selected labroids. TMC, Chorleywood.
- . 2004. Basslets, hamlets, and their relatives: a comprehensive guide to selected Serranidae and Plesiopidae. TMC, Chorleywood.
- Kuiter, R. H., and H. Debelius. 2001. Surgeonfishes, rabbitfishes, and their relatives. TMC, Chorleywood.
- La Monte, F. 1952. Marine game fishes of the world. Doubleday, New York.
- Larson, H. K. 2001. A revision of the gobiid fish genus *Mugilogobius* (Teleostei: Gobioidi), and its systematic placement. Records of the Western Australian Museum 62.
- Leibel, W. S. 1984. Heckel's thread-finned acara: *Acarichthys heckelii* (Mueller and Troschel 1848). Freshwater and Marine Aquarium 1984:15–19, 78–86.
- . 2002. The true basketmouth cichlids of the genus *Acaronia* (Myers 1940). Cichlid News 11:6–17.
- Lewis, D., P. Reinthal, and J. Trendall. 1986. A guide to the fishes of Lake Malawi National Park. Creda, Gland.
- Loiselle, P. V. 1985. The cichlid aquarium. Tetra, New Jersey.
- Losey, G. S., Jr. 1976. The significance of coloration in fishes of the genus *Hypsoblennius* Gill. Bulletin of the Southern California Academy of Sciences 75:183–198.
- Lowe-McConnell, R. H. 1969. The cichlid fishes of Guyana, South America, with notes on their ecology and breeding behavior. Zoological Journal of the Linnean Society 48:255–302.
- Maitland, P. S. 2000. Guide to freshwater fish of Britain and Europe. Octopus, London.
- Malabarba, L. R., M. C. Malabarba, and R. E. Reis. 2015. Descriptions of five new species of the Neotropical cichlid genus *Gymnogeophagus* Miranda Ribeiro, 1918 (Teleostei: Cichliformes) from the Rio Uruguay drainage. Neotropical Ichthyology 13:637–662.
- Masuda, H., K. Amaoka, C. Araga, T. Uyeno, and T. Yoshino. 1984. The fishes of the Japanese archipelago. Tokai University Press, Tokyo.
- McGinnis, S. M., and D. Alcorn. 2006. Field guide to the freshwater fishes of California. University of California Press, Los Angeles.
- Mercy, T. V. A., A. Gopalakrishnan, D. Kapoor, and W. S. Lakra. 2007. Ornamental fishes of the Western Ghats of India. National Bureau of Fish Genetic Resources, Lucknow.
- Michael, S. W. 2004. Angelfishes and butterflyfishes: plus ten more aquarium fish families with expert captive care advice for the marine aquarist. TFH, Neptune City, NJ.
- Michael, S. W. 2004. Basslets, dottybacks, and hawkfishes: plus seven more aquarium fish families with expert captive care advice for the marine aquarist. TFH, Neptune City, NJ.
- . 2008. Damselfishes and anemonefishes: the complete illustrated guide to their identification, behaviors, and captive care. TFH, Neptune City, NJ.
- . 2009. Wrasses and parrotfishes: the complete illustrated guide to their identification, behaviors, and captive care. TFH, Neptune City, NJ.
- Miller, R. G. 1993. A history and atlas of the fishes of the Antarctic Ocean. Foresta Institute for Ocean and Mountain Studies, Carson City, NV.
- Miller, R. J. 1964. Social behavior in the blue gourami *Trichogaster trichopterus* (Pisces: Belontiidae). Copeia 1964:469–496.
- Miller, R. R., W. L. Minckley, and S. M. Norris. 2005. Freshwater fishes of Mexico. University of Chicago Press, Chicago.
- Mills, D., and G. Vevers. 1990. The Tetra encyclopedia of freshwater tropical aquarium fishes. Tetra, New Jersey.

- Morgans, C. L., and T. J. Ord. 2013. Natural selection in novel environments: predation selects for background matching in the body colour of a land fish. *Animal Behavior* 86:1241–1249.
- Moyer, J. T., R. E. Thresher, and P. L. Colin. 1983. Courtship, spawning, and inferred social organization of American angelfishes (Genera *Pomacanthus*, *Holacanthus*, and *Centropyge*, Pomacanthidae). *Environmental Biology of Fishes* 9:25–39.
- Munday, P. L., T. Kuwamura, and F. J. Kroon. 2010. Bidirectional sex change in marine fishes. Pages 241–272 in K. S. Cole, ed. *Reproduction and sexuality in marine fishes: patterns and processes*. University of California Press, Berkeley.
- Myrberg, A. A., Jr., W. L. Montgomery, and L. Fishelson. 1988. The reproductive behavior of *Acanthurus nigrofuscus* (Forsk.) and other surgeonfishes (Fam. Acanthuridae) off Eilat, Israel (Gulf of Aqaba, Red Sea). *Ethology* 79:31–61.
- Nakajima, J. 2012. Taxonomic study of the *Cobitis striata* complex (Cypriniformes, Cobitidae) in Japan. *Zootaxa* 3586:103–130.
- Neat, F. C., and W. Lengkeek. 2009. Sexual selection in blennies. In R. A. Patzner, E. J. Gonçalves, P. A. Hastings, and B. G. Kapoor, eds. *Biology of blennies*. CRC, Boca Raton, FL.
- Nelson, J. S., T. C. Grande, and M. V. H. Wilson. 2016. *Fishes of the world*. 5th ed. Wiley, Hoboken, NJ.
- Noakes, D. L. G., A. Romero, Y. Zhao, and Y. Zhou, eds. 2010. *Chinese fishes*. Vol. 28. Springer, Dordrecht.
- Nugraha, M. F. I., I. W. Subamia, S. Sudarto, and W. Purbowasito. 2011. Sex determination in Indonesian pufferfish *Tetraodon palembangensis* Bleeker, 1852: implication for aquaculture and conservation. *Indonesian Aquaculture Journal* 6:37–45.
- Okada, Y. 1966. *Fishes of Japan: illustrations and descriptions of fishes of Japan*. Uno Shoten, Tokyo.
- Okeyo, D. O., and W. O. Ojwang. 2015. A photographic guide to freshwater fishes of Kenya. *Seriously Fish*. <https://www.seriouslyfish.com/publications>.
- Ord, T. J., and S. T. Hsieh. 2011. A highly social, land-dwelling fish defends territories in a constantly fluctuating environment. *Ethology* 117:918e927.
- Orme, D. 2011. The caper package: comparative analysis of phylogenetics and evolution in R. <https://cran.r-project.org/web/packages/caper/vignettes/caper.pdf>.
- Page, L. M. 1983. *Handbook of darters*. TFH, Neptune City, NJ.
- Page, L. M., and B. M. Burr. 1991. *A field guide to freshwater fishes of North America north of Mexico*. Houghton Mifflin, Boston.
- . 2011. *A field guide to freshwater fishes of North America north of Mexico*. 2nd ed. Houghton Mifflin, Boston.
- Panhuis, T. M., R. Butlin, M. Zuk, and T. Tregenza. 2001. Sexual selection and speciation. *Trends in Ecology and Evolution* 16:364–371.
- Parenti, L. R. 2008. A phylogenetic analysis and taxonomic revision of ricefishes, *Oryzias* and relatives (Belontiiformes, Adrianichthyidae). *Zoological Journal of the Linnean Society* 154:494–610.
- Parenti, L. R., and J. A. Maciolek. 1993. New sicydiine gobies from Ponape and Palau, Micronesia, with comments on systematics of the subfamily Sicydiinae (Teleostei: Gobiidae). *Bulletin of Marine Science* 53:945–972.
- Pastana, M. N. L., F. C. P. Dagosta, and A. L. H. Esguícero. 2017. A new sexually dichromatic miniature *Hyphessobrycon* (Teleostei: Characiformes: Characidae) from the Rio Formiga, upper Rio Juruena Basin, Mato Grosso, Brazil, with a review of sexual dichromatism in Characiformes. *Journal of Fish Biology* 91:1301–1318.
- Pethiyagoda, R. 1991. *Freshwater fishes of Sri Lanka*. Wildlife Heritage Trust of Sri Lanka.
- Pinter, H. 1984. *Labyrinth fish*. Barron's, Woodbury, NY.
- Pollux, B. J. A., R. W. Meredith, M. S. Springer, T. Garland, and D. N. Reznick. 2014. The evolution of the placenta drives a shift in sexual selection in livebearing fish. *Nature* 513:233–236.
- Prokofiev, A. M. 2010. Morphological classification of loaches (Nemacheilinae). *Journal of Ichthyology* 50:827–913.
- Py-Daniel, R., and C. C. Fernandes. 2005. Dimorfismo sexual em Siluriformes e Gymnotiformes (Ostariophysi) da Amazônia. *Acta Amazonica* 35:97–110.
- Rajan, P. T. 2001. *A field guide to grouper and snapper fishes of Andaman and Nicobar Islands*. Zoological Survey of India, New Dehli.
- Randall, J. E. 1983. *Caribbean reef fishes*. TFH Publications, Hong Kong.
- . 2010. *Shore fishes of Hawai'i*. University of Hawaii Press, Honolulu.
- Randall, J. E., G. R. Allen, and R. C. Steene. 1996. *Fishes of the Great Barrier Reef and Coral Sea*. University of Hawaii Press, Honolulu.
- Rass, T. S. 1970. *Greenlings: taxonomy, biology, interoceanic transplantation*. Israel Program for Scientific Translations, Jerusalem.
- Reid, G. M. 1985. A revision of African species of *Labeo* (Pisces: Cyprinidae) and a re-definition of the genus. Braunschweig, Germany.
- Ričan, O., and S. O. Kullander. 2008. The *Australoheros* (Teleostei: Cichlidae) species of the Uruguay and Paraná River drainages. *Zootaxa* 1724:1–51.
- Ričan, O., L. Piálek, A. Almirón, and J. Casciotta. 2011. Two new species of *Australoheros* (Teleostei: Cichlidae) with notes on diversity of the genus and biogeography of the Río de la Plata basin. *Zootaxa* 2982:1–26.
- Ričan, O., L. Piálek, K. Dragová, and J. Novák. 2016. Diversity and evolution of the Middle American cichlid fishes (Teleostei: Cichlidae) with revised classification. *Vertebrate Zoology* 66:1–102.
- Ritchie, M. G., R. M. Hamill, J. A. Graves, A. E. Magurran, S. A. Webb, and C. M. Garcia. 2007. Sex and differentiation: population genetic divergence and sexual dimorphism in Mexican goodeid fish. *Journal of Evolutionary Biology* 20:2048–2055.
- Roberts, C. D., A. L. Stewart, and C. D. Struthers. 2015. *The fishes of New Zealand*. Museum of New Zealand, Wellington.
- Roberts, T. R. 1989. *The freshwater fishes of Western Borneo (Kalimantan Barat, Indonesia)*. *Memoirs of the California Academy of Sciences* 14.
- Robertson, D. R. 1983. On the spawning behavior and spawning cycles of eight surgeonfishes (Acanthuridae) from the Indo-Pacific. *Environmental Biology of Fishes* 9:193–223.
- Robertson, D. R., and G. R. Allen. 2015. *Shorefishes of the tropical eastern Pacific: online information system*. Version 2.0. Smithsonian Tropical Research Institute, Balboa. <https://biogeodb.stri.si.edu/sftstep/en/pages/>.
- Robertson, D. R., and S. G. Hoffman. 1977. The roles of female mate choice and predation in the mating systems of some tropical labrid fishes. *Ethology* 45:298–320.
- Robertson, D. R., and J. Van Tassell. 2015. *Shorefishes of the Greater Caribbean: online information system*. Version 1.0. Smithsonian Tropical Research Institute, Balboa. <https://biogeodb.stri.si.edu/caribbean/en/pages/>.
- Robins, R. C., and G. C. Ray. 1986. *A field guide to Atlantic coast fishes of North America*. Houghton Mifflin, Boston.

- Romero, A., Y. Zhao, and X. Chen. 2010. The hypogean fishes of China. Pages 211–278 in D. L. G. Noakes, A. Romero, Y. Zhao, and Y. Zhou, eds. Chinese fishes. Springer, Dordrecht.
- Rosenblatt, R. H. 1960. The Atlantic species of the blennioid fish genus *Enneanectes*. Proceedings of the Academy of Natural Sciences of Philadelphia 112:1–23.
- Ross, S. T. 2001. The inland fishes of Mississippi. University Press of Mississippi, Jackson.
- Rüber, L., R. Britz, H. H. Tan, P. K. L. Ng, and R. Zardoya. 2004. Evolution of mouthbrooding and life-history correlates in the fighting fish genus *Betta*. Evolution 58:799–813.
- Sandovy, Y., and A. S. Cornish. 2000. Reef fishes of Hong Kong. Hong Kong University Press, Hong Kong.
- Seriously Fish. <https://www.seriouslyfish.com/>.
- Serov, D. V., V. K. Nezdolij, and D. S. Pavlov. 2006. The freshwater fishes of Central Vietnam. KMK Scientific, Moscow.
- Shen, S., T. Yang, and J. J. Lin. 1986. A review of the blennioid fishes in the waters around Taiwan and its adjacent islands. Taiwan Museum, Taipei.
- Sideleva, V. G. 2003. The endemic fishes of Lake Baikal. Backhuys, Leiden.
- Skelton, P. 2001. A complete guide to the freshwater fishes of southern Africa. Southern Book, Halfway House.
- Smith, C., I. Barber, R. J. Wootton, and L. Chittka. 2004. A receiver bias in the origin of three-spined stickleback mate choice. Proceedings of the Royal Society B 271:949–955.
- Smith-Vaniz, W. F. 1976. The saber-toothed blennies, tribe Nemoiphini (Pisces: Blenniidae). Monographs of Academy of Natural Sciences of Philadelphia, No. 19.
- . 1980. Revision of Western Atlantic species of the blennioid fish genus *Hypsoblennius*. Proceedings of the Academy of Natural Sciences of Philadelphia 132:285–305.
- Sparks, J. S. 2008. Phylogeny of the cichlid subfamily Etroplinae and taxonomic revision of the Malagasy cichlid genus *Paretroplus* (Teleostei: Cichlidae). Bulletin of the American Museum of Natural History 314:1–151.
- Sparks, J. S., and W. L. Smith. 2004. Phylogeny and biogeography of the Malagasy and Australasian rainbowfishes (Teleostei: Melanoetaenioidei): Gondwanan vicariance and evolution in freshwater. Molecular Phylogenetics and Evolution 33:719–734.
- Springer, V. G. 1959. Taxonomy and ecology of the genus *Chasmodes*. Texas Journal of Science 11:321–334.
- . 1962. A review of the blennioid fishes of the genus *Ophioblennius* Gill. Copeia 1962:426–433.
- . 1967. Revision of the circumtropical shorefish genus *Entomacrodus* (Blenniidae: Salariaiinae). Proceedings of the United States National Museum 122:1–150.
- . 1988. The Indo-Pacific blennioid fish *Ecsenius*. Smithsonian Contributions to Zoology 465.
- Springer, V. G., and J. T. Williams. 1994. The Indo-West Pacific blennioid fish genus *Istiblennius* reappraised: a revision of *Istiblennius*, *Blenniella*, and *Paralticus*, new genus. Smithsonian Contributions to Zoology 565.
- Sterba, G. 1973. Freshwater fishes of the world. TFH, Neptune City, NJ.
- Stevenson, D. E. 2002. Systematics and distribution of fishes of the Asian goby genera *Chaenogobius* and *Gymnogobius* (Osteichthyes: Perciformes: Gobiidae), with the description of a new species. Species Diversity 7:251–312.
- Stiassny, M. L. J., and A. Getahun. 2007. An overview of labeonin relationships and the phylogenetic placement of the Afro-Asian genus *Garra* Hamilton, 1922 (Teleostei: Cyprinidae) with the description of five new species of *Garra* from Ethiopia, and a key to all African species. Zoological Journal of the Linnean Society 150:41–83.
- Sung, S., and I. Gonzalez. 2008. *Gymnogeophagus*: the glittering eartheaters of the Far South. Tropical Fish Hobbyist.
- Takahashi, T. 2008. Description of a new cichlid fish species of the genus *Benthochromis* (Perciformes: Cichlidae) from Lake Tanganyika. Journal of Fish Biology 72:603–613.
- Tappin, A. R. 2011. Rainbowfishes: their care and keeping in captivity. Art Publications.
- Teletchea, F., A. Fostier, E. Kamler, J. Gardeur, P. Le Bail, B. Jalabert, and P. Fontaine. 2009. Comparative analysis of reproductive traits in 65 freshwater fish species: application to the domestication of new fish species. Reviews in Fish Biology and Fisheries 19:403–430.
- Thacker, C. E., A. R. Thompson, and D. M. Roje. 2011. Phylogeny and evolution of Indo-Pacific shrimp-associated gobies (Gobiiformes: Gobiidae). Molecular Phylogenetics and Evolution 59:168–176.
- Thomas, C., T. H. Bonner, and B. G. Whiteside. 2007. Freshwater fishes of Texas. Texas A&M University Press, College Station.
- Thomson, D. A., L. T. Findley, and A. N. Kerstitch. 1979. Reef fishes of the Sea of Cortez. Wiley, New York.
- Thresher, R. E. 1982. Courtship and spawning in the emperor angelfish *Pomacanthus imperator*, with comments on reproduction by other pomacanthid fishes. Marine Biology 70:149–156.
- Thresher, R. E., and J. T. Moyer. 1983. Male success, courtship complexity and patterns of sexual selection in three congeneric species of sexually monochromatic and dichromatic damselfishes (Pisces: Pomacentridae). Animal Behavior 31:113–127.
- Vagelli, A. A. 2013. Ephemeral sexual dichromatism in *Quinca mirifica* (Teleostei, Apogonidae), a black apogonid with solitary behavior. Aqua 20:1–10.
- Van der Heiden, A. M., and S. M. Pérez. 1992. Secondary sexual dimorphism and dichromatism in Gulf of California flatfishes. Copeia 1992:1077–1080.
- van der Sleen, P., and J. S. Albert, eds. 2018. Field guide to the fishes of the Amazon, Orinoco, and Guianas. Princeton University Press, Princeton, NJ.
- Vassilev, M., A. Apostolou, B. Velkov, D. Dobrev, and V. Zarev. 2012. Atlas of the gobies (Gobiidae) in Bulgaria. Institute of Biodiversity and Ecosystem Research, Bulgarian Academy of Sciences, Sofia.
- Vidthayanon, C., A. Termvidchakorn, and P. Myint. 2005. Inland fishes of Myanmar. Southeast Asian Fisheries Development Center, Bangkok.
- Vincent, A., I. Ahnesjö, A. Berglund, and G. Rosenqvist. 1992. Pipefishes and seahorses: are they all sex role reversed? Trends in Ecology and Evolution 7:237–241.
- Walker, B. 1974. Sharks and loaches. TFH, Neptune City, NJ.
- Warren, M. L., Jr., and B. M. Burr, eds. 2014. Freshwater fishes of North America. Vol. 1. Petromyzontidae to Catostomidae. Johns Hopkins University Press, Baltimore.
- Watanabe, M. 1960. Cottidae (Pisces). Tokyo News Service, Tokyo.
- Watson, R. E., P. Keith, and G. Marquet. 2001. *Sicyopus (Smilosicyopus) chloe*, a new species of freshwater goby from New Caledonia (Sicydiinae). Cybium 25:41–52.
- Wheeler, A. 1978. Key to the fishes of Northern Europe: a guide to the identification of more than 350 species. Frederick Warne, London.

- Whitehead, P. J. P., M. L. Bauchot, J. C. Hureau, J. Nielsen, and E. Tortonese, eds. 1986. Fishes of the north-eastern Atlantic and the Mediterranean. Vol. I-III. United Nations Educational, Scientific, and Cultural Organization, Paris.
- Williams, J. T. 1983. Taxonomy and ecology of the genus *Chasmodes* (Pisces: Blenniidae) with a discussion of its zoogeography. *Bulletin of the Florida State Museum* 29:65-100.
- Winfield, I. J., and J. S. Nelson, eds. 1991. Cyprinid fishes: systematics, biology and exploitation. Chapman & Hall, London.
- Wischnath, L. 1993. Atlas of the livebearers of the world. TFH, Neptune City, NJ.
- Wootton, R. J. 2015. Mating systems and sexual selection. Pages 201-250 in R. J. Wootton and C. Smith, eds. *Reproductive biology of teleost fishes*. Wiley-Blackwell, Sussex.
- Yokoyama, S. 2008. Evolution of dim-light and color vision pigments. *Annual Review of Genomics and Human Genetics* 9:259-282.
- Yonezawa, T., A. Shinomiya, and H. Motomura. 2010. Freshwater fishes of Yaku-shina Island, Kagoshima Prefecture, southern Japan. Pages 249-261 in H. Motomura and K. Matsuura, eds. *Fishes of Yakushina Island*. National Museum of Nature and Science, Tokyo.
- Yuan, L., and E. Zhang. 2010. Morphological variation in *Acrossocheilus hemispinus* (Teleostei: Cyprinidae: Barbinae), with comments on its taxonomic status. *Zootaxa* 2684:45-56.
- Zhuang, P. 2014. Native and exotic fishes of the Middle and Lower Yangtze River. Shanghai Scientific and Technical, Shanghai.

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“Gizzard Shad (*Dorosoma Cepedianum*). We gave a short notice of this species in the ‘Geology of New Jersey,’ page 822, which we will quote and speak of more particularly. ‘Occasionally the “gizzard shad” is carried by a freshet into inland streams usually having very small outlets, and thus imprisoned they thrive very well. A pond near Trenton was, in 1857, stocked with them, and is now full of specimens, some weighing five pounds apiece.’ Besides this pond spoken of we know of one or two creeks that are annually visited by a few of these herring, and have occasionally seen several bushels hauled from the deep holes in the creeks they had entered.” From “Notes on Fresh-Water Fishes of New Jersey” by Charles C. Abbott (*The American Naturalist*, 1870, 4:99-117).